

State of the Science on Coastal Blue Carbon A Summary for Policy Makers

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Executive Summary

Coastal habitats store large amounts of carbon in their living vegetation and soils. When these habitats are converted to other uses, this stored carbon—increasingly known as coastal blue carbon—can be released in the form of greenhouse gases. For this reason, carbon storage should be considered in habitat management decisions. One tool for preserving this critical ecosystem service is payments to landowners and managers for coastal blue carbon—if protocols that allow these carbon stores to be traded on carbon markets can be developed.

The natural science of blue carbon is evolving rapidly, and many policy makers remain uncertain about the biophysical potential of these habitats as engines of carbon storage. To better manage the ecosystem services provided by coastal blue carbon, we need a good scientific understanding of

- how coastal habitats sequester and store carbon,
- where on the planet carbon is stored in these habitats,
- how rapidly the habitats are being modified with a risk of carbon release into the atmosphere or water column, and
- the mechanisms and rate of carbon emissions that follow habitat conversion.

This report examines the current science as it relates to these topics. In doing so, it aims to give policy makers a feel for what is known and unknown about coastal blue carbon. Because of the high intensity of work in this field and variations in research methodologies, additional findings may be available in the near future.

Briefly, what scientists know now is that blue carbon sequestration and storage involves three components. The first is the annual sequestration rate, which in a mature system is the yearly flux of organic material transferred into anaerobic soils, where it cannot undergo oxidation to carbon dioxide (CO_2) and be released to the atmosphere. The second component is the amount of carbon stored in biomass, both above- and belowground. The third and largest component is the total carbon stock stored in soils as a result of prior sequestration. The total carbon stock integrates the complete column of organic soil lying beneath coastal habitats. This stock is a function of the soil carbon density and the depth of the rich, organic soils beneath these ecosystems. Scientists know more about the former than the latter. Generally, total carbon storage estimates are available for at least the first meter of soil—the depth at which carbon is most susceptible to release.

Annual Carbon Sequestration Rates

- Annual carbon sequestration rates have been calculated for 39 mangrove sites. Values range from 0.126 to 23.98 megagrams¹ of carbon dioxide equivalent per hectare per year (Mg CO₂e/ha/yr). Most estimates fall below 7 Mg CO₂e/ha/yr.
- Annual carbon sequestration data are available for 122 coastal marshes. Values range from 0.01 to 62.81 Mg CO₂e/ha/yr. Most estimates fall below 8 Mg CO₂e/ha/yr.
- Annual carbon sequestration data are available for 377 seagrass sites. Values range from -77 to 85 Mg CO₂e/ha/yr. Most estimates fall below 7 Mg CO₂e/ha/yr. A large number of estimates show annual net losses of carbon.

Carbon Stored in Biomass

- Biomass carbon content data are available for 32 mangrove sites. Values range from 25 to 2,254 Mg CO₂e/ha. Most estimates fall between 300 and 1,000 Mg CO₂e/ha.
- Biomass carbon content data are available for 6 salt marsh sites. Values range from 5.1 to 18.3 Mg CO₂e/ha.
- Biomass carbon content data are available for 160 seagrass sites. Values range from 0 to 13 Mg CO₂e/ha.

Soil Carbon Stocks

- Estimates of the soil carbon stock of the first meter of mangrove soils are available for 62 sites. Values range from 570 to 4,712 Mg CO₂e/ha. Most estimates fall between 800 and 3,000 Mg CO₂e/ha.
- Estimates of the soil carbon stock of the first meter of salt marsh soils are available for 126 coastal marshes.

1. One megagram (Mg) = 1 tonne = 1 metric ton = 1,000 kg.

Values range from 174 to 6,967 Mg CO₂e/ha. Most estimates fall between 900 and 1,700 Mg CO₂e/ha.

• Estimates of the total soil carbon stock of the first meter of seagrass soils are available for only 10 sites, all in the Mediterranean. Values range from 880 to more than 6,000 Mg CO₂e/ha.

Geographic Representation

Scientific information about blue carbon habitats is not representative of these habitats worldwide. Understanding of the areal extent of blue carbon habitats and their rate of loss is also not uniform across the globe. In general, habitat extent in the developed world is well known, and habitat extent in Asia is less known. Comparatively little is known about the extent and rates of loss of African coastal habitats.

Mangroves. Understanding of carbon sequestration and storage is geographically uneven. In the United States, most studies are of Florida. Globally, most studies are of Asia and Oceania. The United Nations Food and Agricultural Organization (FAO) amasses mangrove habitat data from around the world. Those data suggest that mangrove habitat loss is highest in absolute terms in Asia, followed by the Americas and Africa.

Salt marshes. Scientific knowledge of carbon sequestration and storage is greatest for eastern North America. Little information is available for the rest of the world. Most is known about the areal extent of salt marsh in North America and Europe, but relatively little is known about the extent of coastal marsh in the rest of the world and very little about rates of salt marsh loss anywhere.

Seagrasses. Understanding of carbon sequestration and biomass storage is greatest for European and North American sites. Little is known about soil carbon associated with seagrasses. Relatively little is known about the extent of seagrass habitat worldwide and even less is known about the loss of that habitat.

Emissions after Conversion

Scientists have amassed little empirical data on the emission of gaseous CO_2 from blue carbon habitats following habitat conversion. Only estimates of actual carbon emissions due to salt marsh conversion are available. To date, all estimates of potential carbon emissions have been estimates bounded by the potential carbon content of soils and biomasses.

1. Some Basics of Blue Carbon Science

Annual carbon sequestration

The annual sequestration rate is the quantity of CO_2 removed from the atmosphere and trapped in natural habitats on an annual basis. By trapping carbon in living tissue, biomass, and soils, these habitats remove carbon from the atmosphere. They can produce greenhouse gases through respiration and through the production of methane in soils (but methane production tends to be low or nonexistent in most marine habitats). They may also export carbon if detritus is removed (e.g., leaves are blown or washed away) or may trap carbon if detritus is captured and incorporated into soils.

Carbon stored in soils

The largest store of carbon in coastal habitats is within soils. To understand the total amount of carbon stored in these soils, scientists must know the carbon density of the soil and the depth of the organic-rich soil layer. The value of the former is typically presented in gC/cm³. Sometimes soil organic carbon is expressed as a percentage of the total soil present. This value can be converted if the bulk density of the soil (g soil/cm³) is known. The depth of the organic-rich soil layer of coastal habitats varies. In some cases, it can be several meters deep, the result of hundreds to thousands of years of growth and accumulation. In this review, we provide data where available on carbon density, soil depth, and when possible, estimates of the total carbon stored in the top meter of a typical hectare of soils.

Carbon stored in biomass

Carbon is stored in aboveground and belowground biomass. Aboveground biomass includes leaves, flowers, stems, branches, and (in the case of mangroves) trunks. Belowground biomass consists mainly of roots and associated flora and fauna. Carbon in biomass is calculated in various ways, as described below.

Areal extent of habitat and habitat conversion

The location, areal extent, and conversion locales and rates of coastal habitats is knowledge critical to identification of opportunities to prevent the emission of trapped carbon from these habitats into the atmosphere. To the extent possible, we document what scientists know about the areal extent of coastal blue-carbon habitats and rates of habitat conversion (often referred to as loss).

Carbon emissions from habitat conversion

Ultimately, the "blue carbon" aspect of coastal habitat hinges on the assumption that large amounts of carbon would be released into the atmosphere if these habitats are disturbed or converted to other uses. Carbon trapped in salt marsh and mangrove habitats is thought to be released directly into the atmosphere through a process of oxidation whereby soil carbon is turned directly into CO_2 when exposed to oxygen. When blue carbon remains under water, the process by which carbon is emitted is less clear. It may be that carbon is mineralized or oxidized in the water column when living biomass dies or when carbon in previously anaerobic soils is exposed to aerobic water. Carbon emitted into the water column could then be released into the atmosphere. Empirical studies that attempt to directly measure the release of carbon into the atmosphere following conversion of blue carbon habitats do not appear to exist.

2. Mangroves

Mangroves are a type of wetland forest with anaerobic sediments. Because the presence of water diminishes oxygen availability, the organic carbon stored in mangrove soils doesn't decompose to atmospheric CO_2 . Once these organic-rich soils are exposed to air, decomposition and production of CO_2 begins. Each gram of organic carbon, stored either as biomass or soil, represents 3.67 CO_2 equivalents.² Many mangrove forests have thousands of years' worth of carbon sequestered beneath them [1]. The depth of these carbon-rich soils varies according to the local geomorphology. For example, mangroves in estuaries tend to have greater depths of organic soils than oceanic mangroves, which have a hard sandy or rocky substrate [2].

Annual sequestration rates of mangrove forests

Two major reviews of carbon sequestration in mangrove habitats have been published. The first, Cebrian (2002) [3], presents data from 8 published studies at 10 sites. It calculates the annual accumulation of carbon or sequestration rate

^{2.} The atomic mass of carbon is 12g/mol. The atomic mass of CO_2 is 44g/mol. Every 12g of organic carbon is equal to 44g of CO_2 equivalents.

using available measurements of community production and respiration. The second important review, by Chmura and colleagues [4], presents data from 5 published studies as well as some unpublished data at 27 sites. It examines organic carbon density within salt marsh and mangrove soils. Combining these density values with annual sediment accretion rates, Chmura and colleagues [4] calculate annual carbon sequestration rates.

This report includes one additional study, by Fujimoto and colleagues [5], that was not included in either of these reviews. Although additional reviews of carbon sequestration in mangrove habitats exist [6-11], they report only summarized data, not individual measurements. Figure 1 displays the geographic representation of the available data. Table A-1 in Appendix A provides a comprehensive summary of the data by geographic location.



Figure 1. Location of data on the annual carbon sequestration rate for mangrove forests.

The range of estimates for carbon sequestration at 39 sites is 0.126 to 23.98 Mg CO₂e/ha/yr (1 Mg CO₂e/ha/yr = 1 megagram of CO₂ equivalents per hectare per year). Figure 2 displays the distribution of the rounded annual sequestration rates. The variability it shows may partly be a function of the two methodologies used. The mean of the values from Cebrian (2002) is 2.87 (n = 9), and the mean of the values from Chmura is 6.96 (n = 28), with a p < 0.05, a two-sample t-test assuming unequal variances. The difference between the means is still significant at the p < 0.05 level, even when the large outlier value of 23.98 is removed from the calculations. How geography, geology, and methodology influence the estimates of sequestration is unclear.



Figure 2. Number of observations for sequestration rates available in the scientific literature.

Soil carbon stocks of mangrove forests

As Figure 3 shows, the organic-rich soils underlying mangrove forests can range in depth from less than a meter [2, 12] to over 10 meters [1]. This report presents 30 observations from four published studies [1, 2, 5, 12]; 25 of the observations are derived from Donato and colleagues [2].



Figure 3. Number of observations for depth data for the organic soil layer beneath mangroves.

Figure 4 presents the geographic representation of the available data on the depth of organic soils beneath mangrove forests.



Figure 4. Location of available mangrove soil depth data.

In assessing estimates of the soil carbon stocks of mangrove forests, there are two key papers to consider. The first is the review by Chmura and colleagues [4], which presents the carbon density of soils. Its data are derived from six published studies and one unpublished source, and it covers 33 locations. The other major article of interest is Donato and colleagues [2], which examines the carbon content of the soil column of 25 mangrove sites in the Indo-Pacific. The Donato paper [2] reports % organic carbon and bulk density data (g/cm³). We use simple multiplication to calculate the carbon density for each of the 25 sites and include data from Fujimoto and colleagues [5] and Vegas-Vilarrubia and colleagues [12], which are not included by Chmura and colleagues [4]. The Fujimoto paper [5] reports organic carbon (gC/kg) and bulk density (kg/m³) at multiple depth intervals; we use these values to calculate a weighted average of carbon density for the entire soil column. Vegas-Vilarrubia and colleagues [12] present organic carbon (g/kg) and bulk density (g/cm³), which we use to calculate carbon density. Other reports contain summaries of review data [13]. However, they were not included here as they presented no new data.

The soil carbon density data, available from 62 sites, ranges from 0.015 to 0.115 gC/cm³. Figure 5 presents the geographic representation of the data. Figure 6 shows the distribution of the available mangrove soil carbon density data. Carbon stocks for the first meter of soil depth were calculated from the available carbon density data using the following conversion:

$$\frac{gC}{cm^3} * \frac{10^6 cm^3}{1m^3} * \frac{10^4 m^2}{1ha} * \frac{44gCO_2 e}{12gC} * \frac{1Mg}{10^6 g} = \frac{MgCo_2 e}{ha * m}$$

Most mangrove habitats have at least one meter of carbon-rich soil; this top meter is at risk if mangroves are converted. On the basis of data from the literature, we find that estimates of carbon stocks in the first meter of mangrove soils range from 570 to 4,217 Mg CO_2e /ha. Figure 7 presents the distribution of these estimates. Table A-2 in Appendix A presents a geographic summary of these data.





Figure 6. Number of observations for mangrove soil carbon density data.





Figure 7. Number of estimates of carbon contained in the top meter of soil beneath mangrove forests.

Carbon content of mangrove biomass

Carbon is stored in the woody biomass of mangrove forests and is typically reported on a dry weight basis. Total biomass estimates can be converted to the carbon content of biomass by using a factor of 45% per dry mass [10]. Two significant papers present empirical data. The most recent is by Donato and colleagues [2] and provides data from 25 mangrove sites throughout the Indo-Pacific. The other is a review by Twilley [10] that provides data from seven published sources on nine sites. Other review papers exist [11, 14] but do not present original data on the topic and are therefore not included here.

A total of 34 observations for the carbon content of mangrove biomass are identified. They range from 26 to 2,554 Mg CO_2e /ha. Figure 8 presents the distribution of the available data for the carbon content of mangrove biomass. Figure 9 presents the geographic representation of these data. The high variability in the numbers is due to the high variability among mangrove species. The FAO recognizes 71 distinct species as true mangroves [15]. These species can range from the size of a small bush to that of a large tree.



Figure 8. Number of observations for data on the carbon content of mangrove biomass.



Figure 9. Location of carbon content of mangrove forest biomass data.

Mangrove habitat extent and loss rates

Mangroves seem to be disappearing at a relatively high rate [15]. The FAO [15] provides the most recent accurate data for mangrove extent. It documents mangrove areas in 124 countries from 1980 through 2005. The data provided in the FAO report build on the Tropical Forest Resources Assessment 1980 [16-18] and on information provided to the Global Forest Resources Assessment 2000 and FRA 2005 [19, 20]. The report combines an extensive review of the literature with a survey of mangrove countries and leading mangrove expert scientists.

Region	1980 Area (103 ha)	2005 Area (103 ha)	Annual Change 2000–2005 (103 ha)	Annual Change 2000–2005 (%)
Africa	3,670	3,160	-12	-0.36
Asia	7,769	5,858	-61	-1.01
North and Central America	2,951	2,263	-18	-0.77
Oceania	2,181	1,972	-8	-0.39
South America	2,222	1,978	-4	-0.18
WORLD	18.794	15.231	-102	-0.66

Table 1. Regional mangrove loss rates [15].

Table excerpted from FAO (2007) [15].

Emissions from mangrove conversion

No specific observational data are available on the amount of carbon released as CO_2 when mangrove forests are converted to other land uses. Donato and colleagues [2] present estimates based on expert opinion, as does a report from Duke University's Nicholas Institute for Environmental Policy Solutions [21], which estimates these releases on the basis of observed data on area loss and carbon storage, as described above. Other land uses, such as forest, also lack direct carbon emissions data at an aggregated scale and require imputation.

3. Salt Marsh

Salt marshes are intertidal ecosystems found on sheltered coastlines ranging geographically from the sub-arctic to the tropics and occurring most extensively in temperate zones. Salt marshes store carbon in anaerobic sediments where it is not oxidized to CO_2 and therefore is not released to the atmosphere. Intertidal ecosystems, such as salt marshes, are dependent on sediment accretion and rising elevation to compensate for sea level rise. As the anaerobic sediments beneath salt marshes accumulate, so too does the total amount of carbon stored in them. Freshwater wetlands tend to be sources of methane (CH_4) [11], a greenhouse gas 25 times more potent than CO_2 . But the saline environment of salt marshes inhibits the natural creation of methane, making for much lower releases of methane in these habitats. The

result is that salt marshes have a much greater capacity for carbon storage than freshwater wetlands.

Annual sequestration rate of salt marshes

Two major articles examine carbon sequestration in salt marshes. The first, by Cebrian [3], presents data from 10 published studies at 19 sites. It calculates this annual accumulation using available measurements of community production and respiration. The second important article, by Chmura and colleagues [4], presents data from 19 published studies as well as some unpublished data at a total of 96 sites. It summarizes information about organic carbon density within salt marsh and mangrove soils. Combining these density values with annual sediment accretion rates, it calculates annual carbon sequestration rate.

This report includes data from these two articles as well as data from six additional studies [22-27]. Yet other studies exist [7, 11] but present only summarized data, not individual measurements.

Estimates of annual carbon sequestration, from 122 observations, range from 0.01 to 62.81 Mg $CO_2e/ha/yr$. The distribution of the available data is shown in Figure 10. The distribution is wide, though clustered primarily in the range of 1 to 10 Mg $CO_2e/ha/yr$. Figure 11 displays the geographic representation of the available data. It shows that the studies largely draw from observations in the eastern North America and Western Europe. Table A-10 in Appendix A provides a comprehensive summary of the data by geographic location.



Figure 10. Number of observations for data on annual carbon sequestration by salt marshes.

The methodology presented by Cebrian [3], which is based on metabolic principles, yields lower estimates than the sediment accumulation methodology used by Chmura and colleagues [4], a two-sample t-test assuming unequal variance, p = 0.05. The average of the estimates from Cebrian (0.13 Mg CO₂e/ha/yr, n = 19) is lower than the average of the estimates by Chmura (8.42 Mg CO₂e/ha/yr, n = 96).





Soil carbon stocks of salt marshes

The rich peaty organic soils underlying salt marshes can range in depth from less than a half a meter [23] to over 7 meters [26, 28], as shown in Figure 12. Brevik and Homburg (2004) [26] note depths of up to 17 meters. However, because their observations were from a coastal wetland system, including a lagoon, only the average depth of 4.57 meters is used here. Drexler and colleagues [28] report a depth of 7.7 meters of salt marsh peat on an island in the Sacramento in the San Joaquin Delta in California. A total of 16 observations are reported here. In some cases, the depths reported are merely the depths that were sampled. The values therefore do not indicate the full depths of the carbon-rich sediments.



Figure 12. Number of observations of data on the depth of organic soils beneath salt marshes.

The leading review article on the carbon density of salt marsh soils is from Chmura and colleagues [4]. The Chmura article presents carbon density data for 106 sites from 20 published papers and several unpublished sources. This report presents these data and data from several other primary sources. Hussein and colleagues [23] present data on soil organic carbon (g/kg) and bulk density (Mg/m³) at various depth increments from nine sampling sites in Dorchester County, Maryland. This report determines the average carbon density for the soil column by calculating a weighted

average of the different depth intervals. Choi and Wang [29] report carbon storage in the top 84 cm of soil beneath a vegetative marsh sequence in St. Marks National Wildlife Refuge in Florida. This report calculates the average carbon density from these available data using average bulk density and percentage soil organic carbon as reported by Johnson and colleagues [24]. Choi and Wang et al. (2004) [25] report a range of carbon density in mmolC/cm³. This report calculates the average of the range and converts it to gC/cm³. Brevik and Homburg [26] also report salt marsh soil carbon density. Craft and colleagues [30] report soil carbon density for two depth intervals at five sites in North Carolina. This report calculates a weighted average of the carbon density by depth at each site.

Estimates of soil carbon stock, from 126 observations, range from 0.009 to 0.190 gC/cm³, though most estimates are found in the lower end of that range. Figure 13 displays the distribution of the data, and Figure 14 displays the geo-graphic representation of the data, again reflecting mostly eastern North America and western Europe.



Figure 13. Number of observations of salt marsh soil carbon density data.



Figure 14. Location of original data on salt marsh soil carbon density.

Carbon stocks for the first meter of soil depth were calculated from the available carbon density data using the conversion presented in the mangrove section above. Carbon stocks in the first meter depth of salt marsh soils ranges from

174 to 6,967 Mg CO_2e /ha. Figure 15 presents the distribution of these estimates. The geographic summary of this data is provided in Table A-11 of Appendix A.



Figure 15. Number of observations of carbon stored in the top meter of salt marsh soil estimates.

Carbon content of salt marsh biomass

Numerous studies dating back to the 1970s measure both above- and belowground biomass of salt marshes. However, data on the carbon content of salt marsh biomass is scarce. One key study by Hemminga and colleagues [31] explores the carbon content of both above- and belowground biomass for salt marshes throughout the growing season. This study shows lower carbon content in salt marsh roots (below ground) than in shoots (above ground) in the beginning of the growing season. However, later in the season the carbon content of both roots and shoots equilibrated at 40% dry weight. We use this factor to convert some of the available peak-season dry-weight biomass data to carbon [32-35]. Biomass estimates vary even within a given species at a given site. For example, Teal and Howes [35] present data for both the tall and short forms of *Spartina alterniflora* at a marsh in Massachusetts. The mean biomass for the tall salt marsh is more than 1,000 g/m² whereas that of the short form is less than 500 g/m². Given the likelihood of variability in carbon content, we transformed only the data from five studies by the 40% carbon content factor in the Hemminga study [31]. We cannot estimate biomass for other salt marsh species because conversion factors, like that given by Hemminga and colleagues [31], are unavailable. Another key study is that of Craft et al. (1988), which specifically examined the carbon content of belowground biomass.

We provide six observations of aboveground biomass from five studies ranging from 5.1 to 18.3 Mg CO_2e/ha . We also present seven observations of belowground biomass from three studies ranging from 3.4 to 51.2 CO_2e/ha . Figure 16 presents the distribution of this data. Figure 17 presents the geographic representation of the data.





Figure 17. Location of data on carbon content of salt marsh biomass.



Salt marsh habitat extent and loss rate

Data on salt marsh area are readily available for North America and Europe and scarce elsewhere. Only one study, by Yang and Chen [36], calculates salt marsh area in China, and it reports a loss of 1.75 million ha of salt marsh between 1950 and 1995. Bridgham and colleagues [11] estimate a loss of 0.4 million ha of North American salt marsh area over the last 200 years. We found no data to estimate salt marsh area or loss rates in South America or Australia (see Table 2). Estimates of current global salt marsh loss rates are 1% to 2% per year [37, 38].

Region	Area (106 ha)	Reference
Unites States	1.926	[4]
Europe and Scandinavia	0.230	[4]
Canada	0.032	[4]
Tunisia	0.006	[4]
Morocco	0.003	[4]
South Africa	0.017	[4]
China	0.350	[36]
Global	140	[38]
North America	2.2	[11]

Table 2. Salt marsh area.

Emissions following salt marsh conversion

Empirical estimates of how much carbon is released to the atmosphere following salt marsh conversion are unavailable. Scientists have attempted to estimate possible carbon releases by examining the subsidence of salt marsh soils following drainage. For centuries, salt marshes subsided after being drained for agricultural purposes. Two types of subsidence occur in marshes. The first, primary subsidence, is a physical change in elevation caused by soil compaction. The mass of the drained soils compacts the soil particles and therefore lowers the elevation. Secondary subsidence is a chemical process whereby exposure of the organic soils of drained marshes to the atmosphere results in oxidation of the soils' organic carbon to CO₂. Much of the published work examining secondary subsidence is focused on freshwater marshes [39, 40]. Data on salt marsh secondary subsidence comes from several sources. Armentano and Menges [40] review available studies of wetland subsidence and soil carbon density from around the world. However, the majority of these studies cover bogs and other fresh water peatlands. The data from the southeastern United States in Armentano and Menges are derived from studies of salt marsh areas. These data are discussed here. Armentano and Menges report a regional mean carbon release rate of 35.64 Mg CO₂e/ha/yr for southeastern U.S. wetlands converted to cropland beginning in 1800.

Drexler and colleagues [28] examine peat loss in the Sacramento in California's San Joaquin Delta. Six of the eight drained wetland agriculture sites they considered were freshwater sites. The Drexler paper reports a total loss of carbon from drained wetlands ranging from 10,633 to 20,900 Mg CO_2e/ha over a period of 125 years. We convert this loss to an annual loss rate ranging from 85 to 167 Mg $CO_2e/ha/yr$.

Ewing and Vepraskas [41] present data on secondary subsidence of a salt marsh converted to agriculture. Their paper examines areas of North Carolina's Juniper Bay that were converted to agriculture. We use the bulk density and percentage carbon data to calculate soil carbon density. We then multiply this density by the secondary subsidence rates to calculate an organic carbon loss rate ranging from 31 to 59 Mg $CO_2e/ha/yr$.

4. Seagrass

Seagrass meadows are communities of underwater-flowering plants found in coastal waters of all continents except Antarctica. Seagrass meadows store relatively small amounts of carbon in aboveground biomass. However, belowground biomass, in the form of large long-lived root structures, stores the majority of carbon below ground. These root structures accumulate large stores of carbon through the formation of "mattes" beneath seagrass meadows. Theses mattes accrete vertically over time, raising the seagrass meadow toward the surface of the water.

Annual carbon sequestration rates of seagrasses

There are two ways to calculate annual carbon sequestration for seagrasses. The first is through the metabolic pathway, which compares annual primary production and community respiration. This method is used in the two review papers discussed below [3, 42]. The second method examines the sedimentary record using radiocarbon dating. Dating the various depths of accumulated sediments allows for calculation of an average annual sequestration rate [43, 44].

Duarte and colleagues [42] use estimates of net production in their review of seagrass sequestration rates and present 358 data points from 34 published studies and at least five unpublished sources.³ Another review, by Cebrian [3],

 $\frac{mmolO_2}{m^2d} * \frac{mmolCO_2}{mmolO_2} * \frac{10^4 m^2}{1ha} * \frac{365.25d}{1yr} * \frac{1molCO_2}{10^3 mmolCO_2} * \frac{44g}{1molCO_2} * \frac{1Mg}{10^6 g} = \frac{MgCO_2}{ha^* yr}$

^{3.} Duarte presents net community production in units of mmol $O_2 m^2 d^{-1}$. We convert these values to Mg $CO_2 e/ha/yr$ using the following calculation:

calculates this annual accumulation or sequestration rate using available measurements of community production and respiration. Cebrian [3] reports a total of 10 estimates of carbon sequestration in seagrasses from six published studies.

The final two papers of interest on this topic are the studies conducted by Mateo and colleagues [43] and Romero and colleagues [44]. The Mateo paper [43] examined seven seagrass sites in the Mediterranean. The Romero paper [44] examined two of those same sites and used a shipwreck site to date the sediments. Mateo and colleagues [43] used radiocarbon methods to date the sedimentary record. Sediment cores were taken from the mattes under these seagrass sites and dated. These dates were then used to calculate an average annual carbon sequestration rate.

We report a total of 377 estimates ranging from -76.70 to 85.44 Mg $CO_2e/ha/yr$. Figure 18 shows the distribution of the data. All the negative values, indicating greater respiration than production, were derived from the Duarte et al. (2010). A one-way ANOVA and Tukey's test was run using Minitab 15 to compare the data sets from the three methodologies (i.e., those of Duarte, Cebrian, and Mateo). No statistically significant ($\alpha = 0.05$) difference was detected among the means of the three data sets. Figure 19 shows the geographic distribution of sites from which the data are collected. A comprehensive summary of the data is presented in Table A-12 of Appendix A.



Figure 18. Number of observations of data on annual carbon sequestration rates of seagrass beds.



Figure 19. Location of estimates of annual carbon sequestration rates for seagrass beds.

Carbon storage under seagrass beds

The largest store of carbon in seagrass systems is contained within the soil. To understand the total amount of carbon stored in the sediments beneath seagrasses, scientists must know the depth of the organic-rich soil layer and the carbon density of the soil, a value typically presented in gC/cm³ but sometimes expressed as a percentage of the total soil present. This value can be converted if the bulk density of the soil (g soil/cm³) is known.

The depth of the "mattes" underneath seagrass meadows is documented only in the Mediterranean. Measured depths range from 1 to 6.5 m. See Table 3.

Table 3. Depth of seagrass mattes.

Location	Number of Observations	Range (m)	Study
Spain/Italy	7	1 – 3.2	[43]
Spain/Italy	2	3.0 – 3.1	[44]
Spain	1	6.5	[45]

Few data on the total carbon stored in seagrass soils are available. We are aware of only three original studies that estimate carbon stored in seagrass soils, and all of them were conducted in the Mediterranean [43-45]. Table 4 presents data on total carbon stored beneath seagrass meadows that integrates both depth and carbon density. Few data on the carbon density of seagrass sediments are available. However, several large data sets on the carbon content of soils as percentage mass of total soil are available [42, 46, 47]. These data sets are not suitable for summary purposes because they do not include the bulk density of seagrass soils, the second component needed to calculate carbon density.

Table 4. Total Carbon stored under seagrass meadows.

Location	Number of Observations	Range (Mg CO₂e/ha)	Study
Spain/Italy	7	1,467 – 5,867	[43]
Spain/Italy	2	880 – 1,760	[44]
Spain	1	6,600	[45]

Carbon content of seagrass biomass

Seagrass meadows sequester carbon in both aboveground and belowground biomass. Duarte and Chiscano (1999) [48] show that aboveground biomass and belowground biomass are roughly equivalent. They use the conversion factor of 0.35 gC/100g dry weight to calculate biomass carbon from dry-weight biomass data. Two review articles compile large data sets of aboveground biomass data [42, 46]. Duarte and colleagues [42] and Kennedy and colleagues [46] present large data sets with 282 unique observations of the carbon content of aboveground seagrass biomass. These data range from 0.01 to 22.75 Mg CO₂e/ha.

Duarte and Chiscano (1999) present aboveground biomass data on 27 species of seagrasses and belowground biomass for 21 species. Although they list the number of observations for each species, they do not identify the locations of the data collections. Therefore, these points are not included in the geographic representation presented in Figure 20 and Table A-13 in Appendix A. These points are included in Figure 21.

The analysis presented by Duarte and Chiscano (1999) reveals that carbon stores in aboveground and belowground biomass are very similar. Therefore, the aboveground biomass numbers presented here may represent only half of the biomass carbon pool.

35 12 10 5 1 6 1 6 5 10 0 2,000 4,000 6,000 0 2,000 4,000 6,000 10,000

Figure 20. Location of seagrass biomass data.





Seagrass habitat extent and loss

Little is known about the areal extent of seagrasses. Table 5 presents the available data on current seagrass area. Waycott and colleagues [49] estimate an annual loss in seagrass habitat of 11,000 ha globally since 1980 and a total loss of 7.2 million ha in the last 100 years.

	Table 5. Available	e data on	seagrass	areal	extent.
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Region	Area (Million ha)	Reference
Mediterranean	2.5 (min)	[50]
Mediterranean	5.0 (max)	[50]
Global	30	[7]
Global	60	[51]
Global	17.7 (min)	[49]
Global	430 (max)	[49]

Appendix A: Summary of Available Data

Table A-1. Available data on annual carbon sequestration rate of mangrove forests.

Location	Number of sites/ observations	Range Mg CO₂e/ha	Average Mg CO₂e/ha	Original references
Florida Keys, USA	15	0.12 – 13.97	5.35	[3, 4, 52-55]
Queensland, Australia	12	0.95 – 12.32	5.76	[3, 4, 56-58]
Estero Pargo, Mexico	4	5.35 – 23.98	11.94	[4, 54]
Hong Kong, China	2	3.75 – 7.8	5.77	[3, 59, 60]
Pohnpei Island, Micronesia	2	1.94 – 3.41	2.68	[5]
Victoria, Australia	1		0.19	[3, 61]
Malaysia	1		5.51	[3, 62]
Puerto Rico	1		1.60	[3, 63]
South Africa	1		4.73	[3, 64]

Table A-2. Available data on total carbon sequestered in the top meter of soils underlying mangrove forests.

Location	Number of sites/	Range Mg CO₂e/ha	Average Mg CO₂e/ha	Original references
	observations			
Orinoco Delta, Venezuela	40*		825	[4, 12]
Florida, USA	12	880 – 2,457	1,588	[4, 53, 54, 65, 66]
Kosrae	7	843 – 2,643	2,085	[2, 4]
Sulawesi	6	2,123 – 2,777	2,487	[2]
Borneo	5	858 – 1,255	1,077	[2]
Үар	5	1,524 – 1,909	1,737	[2]
Boca Chica, Mexico	4	1,723 – 2,127	1,907	[4, 54]
Umengi estuary, South Africa	4	3,850 – 4,217	3,997	[4, 67]
Pohnpei Island, Micronesia	3	1,280 – 2,383	1,814	[5]
Colombia	3	2,126 – 2,603	2,322	[4, 68]
Sundarbans, Bangladesh	2	570 – 587	578	[2]
Java	2	894-1,153	1,024	[2]
Palau	1		1,687	[2]

*This value was calculated reflects the fact that of 227 sites, 49 (22%) represented mangrove habitat, a number decreased to 40 to incorporate a conservative margin of error.

Table A-3. Available data on the depth of organic sediments beneath mangrove forests.

Location	Number of sites/ observations	Range (m)	Average (m)	Original references
Kosrae	11	0.7 – 5.1	2.09	[2, 5, 69]
Sulawesi	6	0.48 – 3	1.22	[2]
Borneo	5	3	3	[2]
Yap	5	1.24 – 2.23	1.62	[2]
Pohnpei Island, Micronesia	4	1.1 – 3.65	2.67	[5, 70]
Sundarbans, Bangladesh	2	3	3	[2]
Java	2	1.42 – 2.81	2.16	[2]
Orinoco Delta, Venezuela	1		0.64	[12]
Pagbilau	1		1.7	[70]
Palau	1		1.17	[2]
Tobacco Range, Belize	1		10	[1]

Location	Number of sites/	Range Mg CO₂e/ha	Average Mg CO₂e/ha	Original references
	observations			
Sulawesi	6	287 – 634	469	[2]
Borneo	5	368 – 946	685	[2]
Үар	5	807 – 2,254	1,550	[2]
Kosrae	4	1,136 – 1,819	1,564	[2]
Sundarbans, Bangladesh	2	338 – 590	464	[2]
Java	2	25.6 – 85	55.3	[2]
Malaysia	2	424 – 473	449	[10]
Mexico	2	198 – 223	210	[10, 71]
Fujijian, China	1		154	[10, 72]
Hainan, China	1		410	[10, 73]
Hong Kong, China	1		213	[10, 74]
Palau	1		681	[2]
Puerto Rico	1		103	[10, 63]
Phuket, Thailand	1		262	[10, 75]

Table A-4. Available data on the carbon content of mangrove biomass.

Table A-5. Mangrove loss rates in Africa [15].

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Madagascar	-3,000	-1
Senegal	-2,400	-2
Guinea-Bissau	-2,200	-1
Sierra Leone	-1,060	-1
Gabon	-588	-0.4
Liberia	-500	-6.1
United Rep. of Tanzania	-440	-0.4
Mozambique	-320	-0.1
Cameroon	-300	-0.1
Ghana	-280	-2.1
Angola	-120	-0.4
Somalia	-100	-1.3
Congo	-70	-0.9
Equatorial Guinea	-60	-0.2
Benin	-40	-3.2
Guinea	-40	n.s.
Dem. Rep. of the Congo	-20	-0.1
Gambia	-20	n.s.
South Africa	-10	-0.3
Côte d'Ivoire	-8	-0.1
Djibouti	0	0
Egypt	0	0
Eritrea	0	0
Kenya	0	0
Mauritania	0	0
Mayotte	0	0
Nigeria	0	0
Seychelles	0	0
Sudan	0	0
Тодо	0	0
Mauritius	6	5.9
Comoros	n.s.	-0.3

Table A-6. Mangrove loss rates in Asia [15].

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Indonesia	-50,000	-1.6
Malaysia	-4,900	-0.8
Philippines	-2,000	-0.8
Myanmar	-1,940	-0.4
Cambodia	-880	-1.2
Thailand	-820	-0.3
Pakistan	-200	-0.1
Viet Nam	-100	-0.1
China	-95	-0.4
India	-40	n.s.
Sri Lanka	-40	-0.4
Iran, Islamic Republic of	-20	-0.1
Bahrain	0	0
Bangladesh	0	0
Brunei Darussalam	0	0
Japan	0	0
Oman	0	0
Qatar	0	0
Saudi Arabia	0	0
Singapore	0	0
Timor-Leste	0	0
Yemen	0	0
United Arab Emirates	20	0.5

Table A-7. Mangrove loss rates in North and Central America [15].

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Mexico	-13,000	-1.5
Honduras	-2,300	-3.1
United States	-1,000	-0.5
Panama	-880	-0.5
Dominican Republic	-520	-2.8
Costa Rica	-160	-0.4
Haiti	-120	-0.8
Belize	-100	-0.1
El Salvador	-100	-0.3
Antigua and Barbuda	-30	-3.8
Cayman Islands	-20	-0.3
Jamaica	-20	-0.2
U.S. Virgin Islands	-10	-5.6
British Virgin Islands	-4	-0.7
Grenada	-3	-1.3
Guadeloupe	-2	-0.1
Barbados	-1	-10.6
Saint Kitts and Nevis	-1	-1.4
Anguilla	0	0
Aruba	0	0
Bahamas	0	0
Bermuda	0	0
Guatemala	0	0
Martinique	0	0

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Montserrat	0	0
Netherlands Antilles	0	0
Nicaragua	0	0
Saint Lucia	0	0
Saint Vincent and the Grenadines	0	0
Trinidad and Tobago	0	0
Turks and Caicos Islands	0	0
Puerto Rico	20	0.2
Cuba	400	0.1
Dominica	n.s.	-2.1

Table A-8, Mangrove	loss rates in	Oceania	[15].
Tuble / Or mangrove	loss rates in	occumu	L

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Papua New Guinea	-6,000	-1.5
Solomon Islands	-760	-1.7
Fiji	-420	-1.1
Australia	-400	n.s.
New Caledonia	-280	-1.6
Samoa	-4	-1.1
American Samoa	-2	-3
Guam	-1	-1.7
Kiribati	0	0
Micronesia (Fed. States of)	0	0
Nauru	0	0
New Zealand	0	0
Niue	0	0
Palau	0	0
Tonga	0	0
Tuvalu	0	0
Vanuatu	0	0
Wallis and Futuna Islands	0	0
Northern Mariana Islands	n.s.	-0.3

Table A-9. Mangrove loss rates in South America [15].

Country	Annual change 2000–2005 (ha)	Annual change 2000–2005 (%)
Colombia	-2,060	-0.6
Venezuela (Bolivarian Rep. of)	-1,500	-0.7
Suriname	-40	n.s.
Brazil	0	0
French Guiana	0	0
Guyana	0	0
Peru	0	0
Ecuador	60	n.s.

Location	Number of observations	Range Mg CO₂e/ha/yr	Average Mg CO₂e/ha/yr	References
Louisiana	34	0.66 – 30.1	10.18	[3, 4, 53, 76-83]
Northeast Canada	31	2.2 – 34.03	8.65	[4, 78, 84, 85]
New England (Connecticut, Massachusetts, and Maine)	20	1.5 –7.5	4.50	[3, 4, 24, 86-90]
Chesapeake Bay, USA	8	1.9 – 12.5	7.12	[3, 4, 23, 91-93]
Europe	7	5.1 - 68.6	31.50	[3, 4, 94-96]
United Kingdom	6	2.8 - 6.8	5.12	[4, 94, 97]
California	5	1.4 – 14.1	6.33	[4, 26, 98, 99]
North Carolina	5	0.8 – 5.3	2.96	[4, 100]
Saint Mark's Wildlife Refuge, Florida	2	1.6 – 6.7	1.88	[4, 25]
Everglades, Florida	2	3.5 – 6.7	5.15	[22]
Rhone Delta, France	1		5.90	[4, 101]
Georgia	1		1.47	[3, 102]
Hong Kong, China	1		6.53	[3, 59]

Table A-10. Available data on annual carbon sequestration rate of salt marshes.

Table A-11. Estimates of carbon stored in the top meter beneath salt marshes using available soil carbon density data.

Location	Number of observations	Range Mg CO₂e/ha	Average Mg CO ₂ e/ ha	References
Northeast Canada	37	660 – 2,680	1,266	[4, 78, 84, 85]
Gulf of Mexico (Louisiana, Texas, and Mississippi)	26	367 – 6,967	1,902	[4, 53, 76-80]
New England (Connecticut, Massachusetts, and Maine)	20	733 – 2,200	1,342	[4, 24, 86-89]
Chesapeake Bay, Maryland	12	902 – 1,613	1,191	[4, 23, 91]
California	6	330 – 1,588	863	[4, 26, 98, 99]
North Carolina	6	174 – 2,038	1,159	[4, 30]
United Kingdom	6	990 – 1,503	1,332	[4, 94, 97]
Florida	5	567 – 1,694	1,020	[4, 25, 29]
Netherlands	4	733 – 1,503	1,201	[4, 94, 95]
Denmark	2	770 – 990	880	[4, 103]
Rhone Delta, France	1		2,677	[4, 101]
British Colombia, Canada	1		623	[4]

Table A-12. Annual carbon sequestration rate data for seagrass meadows worldwide.

Location	Number of sites	Range Mg CO ₂ e/(ha*y)	Average Mg CO ₂ e/	Original references
			(ha*y)	
Florida Keys	84	-9.4 - 50.22	2.83	[3, 42, 104-108]
Spain	57	-11.48 – 21.99	2.87	[3, 42-44, 109-112]
Redfish Bay, Texas, USA	36	-46.96 – 26.62	-6.77	[42, 113]
Northwest Florida	29	-9.97 – 16.97	1.94	[42, 114-116]
Denmark	23	-18.13 – 29.79	0.11	[42, 117]
France	21	-33.63 - 85.44	10.43	[42, 43, 118-121]
Mexico	21	-76.7 – 42.01	8.54	[42, 122]
Chesapeake Bay, USA	19	-12.33 – 62.13	13.81	[42, 104, 123, 124]
Alabama	13	-1.39 – 53.54	25.8	[42, 125]
Laguna Madre, Texas	13	-11.05 – 62.78	10.61	[42, 126, 127]
Gulf of Carpentaria, Australia	12	-2.68 – 22.77	9.37	[42, 128]
Indonesia	12	-52.54 – 2.83	-21.19	[42, 129]
Greece	6	4.35 – 10.26	6.33	[42, 130]
Philippines	4	-1.25 – 22.53	7.81	[42, 131]
East Coast of Australia	3	-14.55 – -5.2	-9.31	[42, 132]
Beaufort North Carolina	3	0.38 – 2.4	1.24	[3, 108]
New England (NH, MA) USA	3	-1 – 1.51	0	[42, 124]
The Bahamas	3	26.79 – 35.62	30.62	[42, 133]
Norway	2	-0.02 - 1.6	0.79	[42, 134]

Location	Number of sites	Range Mg CO₂e/(ha*y)	Average Mg CO₂e/ (ha*y)	Original references
Puerto Rico	2	25.11 – 75.33	50.22	[42, 135]
India	1		77.81	[42, 136]
South Africa	1		13.4	[3, 137]
Wadden Sea, Germany	1		1.21	[42, 138]
The Netherlands	1		1.87495	[3, 139]
Padilla Bay, Washington, USA	1		-2.01	[42, 124]

Table A-13. Distribution of seagrass biomass sites/observations.

Location	Number of sites	Range Mg CO₂e/ha	Average Mg CO ₂ e/ ha	Original references
Florida Keys	62	0.21 – 3.67	0.62	[42, 46, 105, 115, 140-143]
Spain	36	1.87 – 15.58	7.91	[42, 46, 110, 111, 144]
Gulf Coast of Florida	35	0.01 – 5.00	1.04	[42, 114-116, 145]
Denmark	23	0.06 – 5.90	1.89	[42, 117, 146]
Flores Sea, Indonesia	17	0.26 – 4.91	2.28	[42, 147]
Gulf of Mexico (AL, MI)	15	0.06 – 2.12	0.41	[42, 125, 148, 149]
France	13	0.14 – 2.19	0.84	[42, 106, 118, 119]
Southwest Australia	12	0.06 – 3.32	1.22	[42, 128]
Spermonde Archipelago, Indonesia	12	0.21 – 3.05	1.26	[42, 129]
Ebro Delta, Spain	12	0.02 – 3.18	1.52	[42, 46, 109]
Chesapeake Bay (VA and MD)	11	0.29 – 1.42	0.83	[42, 123]
Greece	9	1.1 – 8.42	3.47	[42, 46, 130, 150]
Philippines	9	0.08 - 3.00	1.44	[42, 46, 131, 151]
Portugal	7	8.0 – 13.20	11.10	[42, 138]
Northeast Australia	6	0.20 – 22.75	7.18	[46]
Alicante, Spain	3	6.34 – 10.75	8.78	[46, 150]
Italy	2	0.62 – 3.68	2.15	[42, 46, 130, 150]
Norway	2	1.75 – 1.84	1.79	[42, 134]
Bay of Cadiz, Spain	2	0.65 – 1.21	0.93	[46]
Thailand	2	0.52 – 0.61	0.56	[46, 152]
Wadden Sea, Germany	1		10.67	[42, 138]
North Carolina, USA	1		1.30	[42, 153]
Rhode Island, USA	1		0.82	[42, 154]

Appendix B: Bibliography by Species

Mangrove References

- Alongi, D.M., F. Tirendi, P. Dixon, L.A. Trott, and G.J. Brunskill. 1999. Sources, sinks, and export of organic carbon fluzes in adjacent tropical nearshore sediments. *Marine Ecological Progress Series* 56:134–144.
- Bouillion, S., V.H. Rivera-Monroy, R.R. Twilley, and J.G. Kairo. 2009. Mangroves. Gland, Switzerland: IUCN.
- Bouillion, S., A.V. Borges, E. Castaneda-Moya, K. Diele, T. Dittmar, N.C. Duke, E. Kristensen, S.Y. Lee, C. Marchand, J.J. Middleburg, V.H. Rivera-Monroy, T.J.I. Smith, and R.R. Twilley. 2008. Mangrove production and carbon sinks: A revision of global budget estimates. *Global Biogeochemical Cycles* 22:GB2013.
- Bridgham, S.D., J.P. Megonigal, J.K. Keller, N.B. Bliss, and C. Trettin. 2006. The carbon balance of North American wetlands. *Wetlands* 26(4):889–916.
- Brunskill, G.J., J. Zagorskis, and J. Pfitzner. 2002. Carbon burial rates in sediments and a carbon mass balance, of the Herbert River region of the Great Barrier Reef continental shelf, north Queensland, Australia. *Estuarine Coastal and Shelf Science* 54:677–700.
- Cahoon, D.R. and J.C. Lynch. 1997. Vertical accretion and shallow subsidence in a mangrove forest of southwestern Florida. *Mangroves and Salt Marshes* 1:173–186.
- Callaway, J.C., R.D. DeLuane, and W.H.J. Patrick. 1997. Sediment accretion rates from four coastal wetlands along the Gulf of Mexico. *Journal of Coastal Research* 13:181–191.
- Cardona, P. and L. Botero. 1998. Soil characteristics and vegetation structure in a heavily deteriorated mangrove forest in the Caribbean coast of Colombia. *Biotropica* 30:24–34.
- Cebrian, J. 2002. Variability and control of carbon consumption, export, and accumulation in marine communities. *Limnology and Oceanography* 47(1):11–22.
- Chen, R.H. and R.R. Twilley. 1999. A simulation model of organic matter and nutrient accumulation in mangrove wetland soils. *Biogeochemistry* 44(1):93–118.
- Chmura, G.L., S.C. Anisfeld, D.R. Cahoon, and J.C. Lynch. 2003. Global carbon sequestration in tidal, saline wetland soils. *Global Biogeochemical Cycles* 17(4):1–12.
- Christensen, B. 1978. Biomass and primary production of Rhizophora apiculata B1. in a mangrove forest in southern Thailand. *Aquatic Botany* 4:43–52.
- Day, J., W. Conner, F. Ley-Lou, R. Day, and S. Machado. 1987. The productivity and composition of mangrove forests, Laguna de Terminos, Mexico. *Aquatic Botany* 27:267–284.
- Donato, D., J.B. Kauffman, D. Murdiyarso, S. Kurnianto, M. Stidham, and M. Kanninen. 2011. Mangroves among the most carbon-rich forests in the tropics. *Nature Geoscience Online*: April 3.
- Duarte, C.M., J.J. Middleburg, and N. Caraco. 2005. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2:1–8.
- Emmerson, W.D. and L.E. McGwynne. 1992. Feeding and assimilation of mangrove leaves by the crab Sesarma meinerti de Man in relation to leaf-litter production in Mgazana, a warm-temperate southern African mangrove swamp. *Journal of Experimental Marine Biology and Ecology* 157(1):41–53.
- FAO (Food and Agriculture Organization). 2001. Global forest resources assessment 2000: Main report. Rome: FAO. http://www.fao.org/forestry/fo/fra/main/index.jsp.
- ———. 2006. Global forest resources assessment 2005: Progress toward sustainable forest management. Rome: FAO. http://www.fao.org/forestrysite/fra200.
- -----. 2007. The World's Mangroves 1980-2005. Rome: FAO.
- Fujimoto, K. and T. Miyagi. 1993. Development process of tidal-flat type mangrove habitats and their zonation in the Pacific Ocean. *Vegetatio* 106:137–146.
- Fujimoto, K., T. Miyagi, T. Kikuchi, and T. Kawana. 1996. Mangrove habitat formation and response to holocene sealevel changes on Krosae Island, Micronesia. *Mangroves and Salt Marshes* 1:47–57.

- Fujimoto, K., A. Imaya, R. Tabuchi, S. Kuramoto, H. Utsugi, and T. Murofushi. 1999. Belowground carbon storage of Micronesian mangrove forests. *Ecological Research* 14:409–413.
- Golley, F., H.T. Odum, and R. Wilson. 1962. The structure and metabolism of a Puerto Rican red mangrove forest in May. *Ecology* 43:8–19.
- Jennerjahn, T.C. and V. Ittekkot. 2002. Relevance of mangroves for the production and deposition of organic matter along tropical continental margins. *Naturwissenschaften* 89:23–30.
- Lee, S. 1990a. Net aerial primary productivity, litter production and decomposition of the reed Phragmites communis in a nature reserve in Hong Kong: Management implications. *Marine Ecology Progress Series* 66:161–173.
- Lee, S.Y. 1990b. Primary productivity and particulate organic matter flow in an estuarine mangrove-wetland in Hong Kong. *Marine Biology* 106(453–463).
- Li, M.S. and S.Y. Lee. 1998. Carbon dynamics of Deep Bay, eastern Pearl River Estuary, China: A mass balance budget and implications for shorebird conservation. *Marine Ecology Progress Series* 172:73–87.
- Lin, P., C.Y. Lu, G.L. Wang, and H.X. Chen. 1990. Biomass and productivity of Bruguiera sexangu: A mangrove forest in Hainan Island, China. *Journal of Xiamen University* 29:209–213.
- Lin, P., C.Y. Lu, G.H. Lin, R.H. Chen, and L. Su. 1985. The biomass and productivity of Kandelia camel community. *Journal of Xiamen University* 14:508–514.
- Lynch, J.C. 1989. Sedimentation and nutrient accumulation in mangrove ecosystems of the Gulf of Mexico. Lafayette, LA: University of Southwestern Lousiana.
- Macintyre, I.G., M.M. Littler, and D.S. Littler. 1995. Holocene history of Tobacco Range, Belize, Central America. *Atoll Research Bulletin* 430:1–18.
- Murray, B.C., L. Pendleton, W.A. Jenkins, and S.D. Sifleet. 2011. Green payments for blue carbon: Economic incentives for protecting threatened coastal habitats. Durham, NC: Nicholas Institute for Environmental Policy Solutions, Duke University.
- Naidoo, G. 1980. Mangrove soils of the Beachwood area, Durban. Journal of South African Botany 46:293-304.
- Nellemann, C., E. Corcoran, C.M. Duarte, L. Valdes, C. De Young, L. Fonseca, and G. Grimsditch. 2009. Blue Carbon: A Rapid Response Assessment. United Nations Environment Programme. http://www.grida.no.
- Ong, J.E. 1993. Mangroves: A carbon source and sink. Chemosphere 27:1097-1107.
- PWA and SAIC. 2009. Greenhouse gas mitigation typology issues paper. Tidal Wetlands Restoration. PWA REF. 1957 California Climate Action Registry. 69 pp.
- Robertson, A. and P. Daniel. 1989. Decomposition and the annual flux of detritus from fallen timber in tropical mangrove forests. *Limnology and Oceanography* 34:640–647.
- Ross, M.S., M.S. Meeder, J.P. Sah, P.L. Ruiz, and G.J. Telesnicki. 2000. The southeast saline Everglades revisited: 50 years of coastal vegetation change. *Journal of Vegetation Science* 11:101–112.
- Twilley, R., A. Lugo, and C. Patterson-Zuca. 1986. Litter production and turnover in basin mangrove forests in southwest Florida. *Ecology* 67:670–683.
- Twilley, R.R., R.H. Chen, and T. Hargis. 1992. Carbon sinks in mangroves and their implications for carbon budget of tropical ecosystems. *Water, Air, and Soil Pollution* 63:265–288.
- UNEP (United Nations Environment Programme) and FAO. 1981a. Tropical forest resources assessment project: Forest resources of tropical Asia. Rome: UNEP and FAO.
- ———. 1981b. Tropical forest resources assessment project: Forest resources of tropical Africa. Part II. Country briefs. Rome: UNEP and FAO.
- ———. 1981c. Proyecto de evaluacion de los recursos forestales tropicales: Los recursos forestales de la America tropicale. Rome: UNEP and FAO.
- Van der Valk, A. and P. Attiwill. 1986. Decomposition of leaf and root litter of Aviccenia marina at Westernport Bay, Victoria, Autralia. *Aquatic Botany* 18:205–221.

Vegas-Vilarrubia, T., F. Baritto, P. Lopez, G. Melean, M.E. Ponce, L. Mora, and O. Gomez. 2010. Tropical histosols of the lower Orinoco Delta: Features and preliminary quantification of their carbon storage. *Geoderma* 155:280–288.

Salt Marsh References

Adam, P. 2002. Salt marshes in a time of change. Environmental Conservation 29(1):39-61.

- Anisfield, S.C., M.J. Tobin, and G. Benoit. 1999. Sedimentation rates in a flow-restricted and restored salt marshes in Long Island Sound. *Estuaries* 22:231–244.
- Armentano, T.V. and E.S. Menges. 1986. Patterns of change in the carbon balance of organic soil wetlands of the temperate zone. *Journal of Ecology* 74(3):755–774.
- Blum, L. 1993. Spartina alterniflora root dynamics in a Virginia marsh. Marine Ecology Progress Series 102:169–178.
- Brevik, E.C. and J.A. Homburg. 2004. A 5000-year record of carbon sequestration from a coastal lagoon and wetland complex, Southern California, USA. *Catena* 57:221–232.
- Bridgham, S.D., J.P. Megonigal, J.K. Keller, N.B. Bliss, and C.C. Trettin. 2006. The carbon balance of North American wetlands. *Wetlands* 26(4):889–916.
- Bryant, J.C. and R.H. Chabreck. 1998. Effects of impoundment on vertical accretion of coastal marsh. *Estuaries* 21:416–422.
- Buth, G. 1987. Decomposition of roots of three plant communities in a Dutch salt marsh. Aquatic Botany 29:123–128.
- Cahoon, D.R. 1994. Recent accretion in two managed marsh impoundments in coastal Louisiana. Ecological Applications 41:166–176.
- Cahoon, D.R. and J.C. Stevenson. 1986. Production, predation and decomposition in a low-salinity Hibischus marsh. *Ecology* 67:1341–1350.
- Cahoon, D.R. and R.E. Turner. 1989. Accretion and canal impacts in a rapidly subsiding wetland: Feldspar marker horizon technique. *Estuaries* 12:260–268.
- Cahoon, D.R., J.C. Lynch, and A. Powell. 1996. Marsh vertical accretion in a southern California estuary, U.S.A. *Estuarine Coastal and Shelf Science* 43:19–36.
- Callaway, J.C., R.D. DeLuane, and W.H.J. Patrick. 1996. Chernobyl Cs used to determine sediment accretion rates at selected northern European salt marshes. *Limnology and Oceanography* 41:444–450.
- Callaway, J.C., R.D. DeLuane, and W.H.J. Patrick. 1997. Sediment accretion rates from four coastal wetlands along the Gulf of Mexico. *Journal of Coastal Research* 13:181–191.
- Cebrian, J. 2002. Variability and control of carbon consumption, export, and accumulation in marine communities. *Limnology and Oceanography* 47(1):11–22.
- Chmura, G.L. and G. Hung. 2004. Controls on salt marsh accretion: A test in salt marshes of Eastern Canada. *Estuaries* 27(1):70–81.
- Chmura, G.L., S.C. Anisfeld, D.R. Cahoon, and J.C. Lynch. 2003. Global carbon sequestration in tidal, saline wetland soils. *Global Biogeochemical Cycles* 17(4):1–12.
- Choi, Y. and Y. Wang. 2001. Vegetation succession and carbon sequestration in a coastal wetland in northwest Florida: Evidence from carbon isotopes. *Global Biogeochemical Cycles* 15(2):311–319.
- Choi, Y. and Y. Wang. 2004. Dynamics of carbon sequestration in a coastal wetland using radiocarbon measurements. *Global Biogeochemical Cycles* 18:GB4016.
- Connor, R.F., G. Chmura, and C.B. Beecher. 2001. Carbon accumulation in Bay of Fundy salt marshes: Implications for restoration of reclaimed marshes. *Global Biogeochemical Cycles* 15(4):934–954.
- Craft, C.B. and C.J. Richardson. 1998. Recent and long-term organic soil accretion and nutrient accumulation in the Everglades. *Soil Science Society of America Journal* 62:834–843.
- Craft, C.B., S.W. Broome, and E.D. Seneca. 1988. Nitrogen, phosphorus, and organic carbon pools in natural and transplanted marsh soils. *Estuaries* 11(4):272–280.
- Craft, C.B., E.D. Seneca, and S.W. Broome. 1993. Vertical accretion in micro-tidal regularly and irregularly flooded estuarine marshes. *Estuarine Coastal and Shelf Science* 37:371–386.

- Day, J., W. Smith, P. Wagner, and W. Stone. 1973. Community structure and carbon budget of a salt marsh and shallow bay estuarine system in Louisiana. Baton Rouge, LA: Center for Wetland Resources, Lousiana State University.
- Drexler, J.Z., C.S. de Fontaine, and S.J. Deverel. 2009. The legacy of wetland drainage on the remaining peat in the Sacramento-San Jooaquin Delta, California, USA. *Wetlands* 29(1):372–386.
- Duarte, C.M., J.J. Middleburg, and N. Caraco. 2005. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2:1–8.
- Duarte, C.M., W.C. Dennison, R.J.W. Orth, and T.J.B. Carruthers. 2008. The charisma of coastal ecosystems: Addressing the imbalance. *Estuaries and Coasts* 31:233–238.
- Ewing, J.M. and M.J. Vepraskas. 2006. Estimating primary and secondary subsidence in an organic soil 15, 20, and 30 years after drainage. *Wetlands* 26(1):119–130.
- French, J.R. and T. Spencer. 1993. Dynamics of sedimentation in a tide-dominated back barrier salt marsh, Norfolk, UK. Marine Geology 110:315–331.
- Giani, L., K. Dittrich, A. Martsfeld-Hartmann, and G. Peters. 1996. Methanogenesis in saltmarsh soils of the north sea coast of Germany. European Journal of Soil Science 47:175–182.
- Hatton, R.S. 1981. Aspects of marsh accretion and geochemistry: Barataria Basin. Baton Rouge, LA: Lousiana State University.
- Hemminga, M.A., A.H.L. Huiskes, M. Steegstra, and J. van Soelen. 1996. Assessment of carbon allocation and biomass production in a natural stand of the salt marsh plant *Spartina anglica* using 13C. *Marine Ecology Progress Series* 130:169–178.
- Hensel, P.F., J.W.J. Day, and D. Pont. 1999. Wetland vertical accretion and soil elevation change in the Rhone River Delta, France: The importance of riverine flooding. *Journal of Coastal Research* 15:668–681.
- Hopkinson, O., J.G. Gosselink, and P. Panardo. 1978. Aboveground production of seven marsh plant species in coastal Lousiana. *Ecology* 59:760–769.
- Howes, B.L., J. Dacey and J.M. Teal. 1985. Annual C mineralization and belowground production of *Spartina alterniflora* in a New England salt marsh. *Ecology* 66:595–605.
- Huang, Y., W. Sun, W. Zhang, Y. Yu, Y. Su, and C. Song. 2009. Marshland conversion to cropland in northeast China from 1950 to 2000 reduced the greenhouse effect. *Global Change Biology* 16(2):680–695.
- Hussein, A.H., M.C. Rabenhorst, and M.L. Tucker. 2004. Modeling of carbon sequestration in coastal marsh soils. *Soil Science Society of America Journal* 68:1786–1795.
- Johnson, B.J., K.A. Moore, C. Lehmann, C. Bohlen, and T.A. Brown. 2007. Middle-to-late Holocene fluctuations of C3 and C4 vegetation in a northern New England salt marsh, Sprague Marsh, Phippsburg, Maine. *Organic Geochemistry* 38:398–403.
- Kearney, M.S. and J.C. Stevenson. 1991. Island land loss and marsh vertical accretion rate evidence for historical sealevel changes in the Chesapeake Bay. *Journal of Coastal Research* 7:403–415.
- Lee, S. 1990. Net aerial primary productivity, litter production and decomposition of the reed Phragmites communis in a nature reserve in Hong Kong: Management implications. *Marine Ecology Progress Series* 66:161–173.
- Markewich, H.W. 1998. Carbon storage and late Holocene chronostratigraphy of a Mississippi River deltaic marsh. U.S. Geological Survey Open File Report 98-36.
- McCaffery, R.J. and J. Thomson. 1980. A record of the accumulation of sediment trace metals in a Connecticut salt marsh. *Advances in Geophysics* 22:165–236.
- Morgan, P.A. and F.T. Short. 2002. Using functional trajectories to track constructed salt marsh development in the Great Bay Estuary, Maine/New Hampshire, U.S.A. *Restoration Ecology* 10(3):461–473.
- Morris, J.T. and B. Haskin. 1990. A 5-year record of aerial primary production and stand characteristics of *Spartina alterniflora*. *Ecology* 71(6):2209–2217.
- Morris, J.T. and A. Jensen. 1998. The carbon balance of grazed and non-grazed *Spartina anglica* salt marshes at Skalligen, Denmark. *Journal of Ecology* 86:229–242.

- Oenema, O. and R.D. DeLuane. 1988. Accretion rates in salt marshes in the Eastern Scheldt, southwest Netherlands. *Estuarine Coastal and Shelf Science* 26:379–394.
- Orson, R.A., R.S. Warren, and W.A. Niering. 1998. Interpreting sea-level rise and rates of vertical marsh accretion in a southern New England tidal salt marsh. *Estuarine Coastal and Shelf Science* 26:419–429.
- Patrick, W.H.J. and R.D. DeLuane. 1990. Subsidence, accretion, and sea-level rise in south San Fransisco Bay marshes. *Limnology and Oceanography* 35:1389–1395.
- Roman, C.T., A. Peck, J.R. Allen, J.W. King, and P.G. Appelby. 1997. Accretion of a New England (U.S.A.) salt marsh in a response to inlet migration, storms and sea-level rise. *Estuarine Coastal and Shelf Science* 45:717–727.
- Teal, J.M. 1962. Energy flow in the salt marsh ecosystem of Georgia. *Ecology* 43:615–624.
- Teal, J.M. and B.L. Howes. 1996. Interannual variability of a salt-marsh ecosystem. *Limnology and Oceanography* 41(4):802–809.
- White, D., T. Weiss, J. Trapani, and L. Thien. 1978. Production and decomposition of the dominant salt marsh plants in Lousiana. *Ecology* 59:751–759.
- Yang, S. and J. Chen. 1995. Coastal salt marshes and mangrove swamps in China. Chinese *Journal of Oceanology and Limnology* 13(4):318–324.
- Yu, O.T. and G.L. Chmura. 2009. Soil carbon may be maintained under grazing in a St. Lawrence Estuary tidal marsh. *Environmental Conservation* 36(4):312–320.

Seagrass References

- Anton, A., J. Cebrian, Duarter, K.L. Heck, Jr., and J. Goff. 2009. Low impact of Hurricane Katrina on seagrass community structure and functioning in the northern Gulf of Mexico. *Bulletin of Marine Science* 85(1):45–59.
- Apostolaki, E.T., M. Holmer, N. Marba, and I. Karakassis. 2010. Metabolic imbalance in coastal vegetated (*Posidonia oceanica*) and unvegetated benthic ecosystems. *Ecosystems* 13:459–471.
- Armitage, A.R. and J.W. Fourqurean. 2009. Stable isotopes reveal complex changes in trophic relationships following nutrient addition in a coastal marine ecosystem. *Estuaries and Coasts* 32:1152–1164.
- Asmus, R.M., M. Sprung, and H. Asmus. 2000. Nutrient fluxes in intertidal communities of a South European lagoon (Ria Formos): Similarities and differences with a northern Wadden Sea bay (Sylt-Romo Bay). *Hydrobiologia* 436:217–235.
- Baird, D. and R.E. Ulanowicz. 1993. Comparative study on the trophic structure, cycling and ecosystem properties of four tidal estuaries. *Marine Ecology Progress Series* 99:221–237.
- Barron, C. and C.M. Duarte. 2009. Dissolved organic matter (DOM) release in a *Posidonia oceanica* meadow. *Marine Ecological Progress Series* 374:75–84.
- Barron, C., N. Marba, J. Terrados, H. Kennedy, and C.M. Duarte. 2004. Community metabolism and carbon budget along a gradient of seagrass (*Cymodocea nodosa*) colonization. *Limnology and Oceanography* 49:1642–1652.
- Bouillon, S. and H.T.S. Boschker. 2006. Bacterial carbon sources in coastal sediments: A cross-system analysis based on stable isotope data of biomarkers. *Biogeosciences* 3:175–185.
- Caffrey, J.M. 2004. Factors controlling net ecosystem metabolism in U.S. estuaries. Estuaries 27:90-101.
- Calleja, M.L., C. Barron, J.A. Hale, T.K. Frazer, and C.M. Duarte. 2006. Light regulation of benthic sulfate reduction rates mediated by seagrass (Thalassia testudinum) metabolism. *Estuaries and Coasts* 29:1255–1264.
- Cebrian, J. 2002. Variability and control of carbon consumption, export, and accumulation in marine communities. *Limnology and Oceanography* 47(1):11–22.
- Cebrian, J., M.F. Pedersen, K. Kroeger, and I. Valiela. 2000. Fate and production of the seagrass *Cymodocea nodosa* in different stages of meadow formation. *Marine Ecology Progress Series* 204:119–130.
- Charpy-Roubaud, C. and A. Sournia. 1990. The comparative estimation of phytoplankton microphytobenthic production in the oceans. *Marine Microbial Food Webs* 4:31–57.
- Duarte, C.M. and C.L. Chiscano. 1999. Seagrass biomass and production: A reassessment. *Aquatic Botany* 65:159–174.

- Duarte, C.M., R. Martinez, and C. Barron. 2002. Biomass, production and rhizome growth near the northern limit of seagrass (Zostera marina) distribution. *Aquatic Botany* 72:183–189.
- Duarte, C.M., J.J. Middleburg, and N. Caraco. 2005. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2:1–8.
- Duarte, C.M., N. Marba, E. Gacia, J.W. Fourqurean, J. Beggins, C. Barron, and E.T. Apostolaki. 2010. Seagrass community metabolism: Assessing the carbon sink capacity of seagrass meadows. *Global Biogeochemical Cycles* 24(GB4032):1–8. Erftemeijer, P.L.A., R. Osinga, and A.E. Mars. 1993. Primary production of seagrass beds in South Sulawesi (Indonesia): A comparison of habitats, methods and species. *Aquatic Botany* 46:67–90.
- Eyre, B.D. and J.P. Ferguson. 2002. Comparison of carbon production and decomposition, benthic nutrient fluxes and denitrification in seagrass, phytoplankton, benthic microalgae- and macroalgae-dominated warm-temperate Australian lagoons. *Marine Ecology Progress Series* 229:43–59.
- Fourqurean, J.W., M.F. Muth, and J.N. Boyer. 2010. Epiphyte loads on seagrasses and microphytobenthos abundance are not reliable indicators of nutrient availability in coastal ecosystems. *Marine Pollution Bulletin* 60:971–983.
- Fourqurean, J.W., A. Willsie, C.D. Rose, and L.M. Rutten. 2001. Spatial and temporal pattern in seagrass community composition and productivity in south Florida. *Marine Biology* 138:341–354.
- Fourqurean, J.W., S.P. Escorcia, W.T. Anderson, and J.C. Zieman. 2005. Spatial and seasonal variability in elemental content, d13C and d15N of Thalassia testudinum from south Florida. *Estuaries* 28(3):447–461.
- Frankingnoulle, M. and J.M. Bouquegneau. 1987. Seasonal variation of the diel carbon budget of a marine macrophyte ecosystem. *Marine Ecology Progress Series* 38:197–199.
- Gacia, E., H. Kennedy, C.M. Duarte, J. Terrados, N. Marba, S. Papadimitriou, and M. Fortes. 2005. Light-dependence of the metabolic balance of a highly productive Philippine seagrass community. *Journal of Experimental Marine Biology and Ecology* 316:55–67.
- Gazeau, F., C.M. Duarte, J.-P. Gattuso, C. Barron, N. Navarro, S. Ruiz, Y.T. Prairie, M. Calleja, B. Delille, M. Frankignoulle, and A.V. Borges. 2005. Whole-system metabolism and CO₂ fluxes in a Mediterranean Bay dominated by seagrass beds (Palma Bay, NW Mediterranean). *Biogeosciences* 2:43–60.
- Heffernan, J.J. and R.A. Gibson. 1983. A comparison of primary production rates in Indian River, Florida seagrass systems. *Florida Science* 48:295–306.
- Herbert, D.A. and J.W. Fourqurean. 2008. Ecosystem structure and function still altered two decades after short-term fertilization of a seagrass meadow. *Ecosystems* 11:688–700.
- Holmer, M., F.O. Andersen, S.L. Nielsen, and H.T.S. Boschker. 2001. The importance of sulfate reduction based mineralization for nutrient regeneration in tropical seagrass sediments. *Aquatic Botany* 71:1–17.
- Holmer, M., C.M. Duarte, H.T.S. Boschker, and C. Barron. 2004. Carbon cycling and bacterial carbon sources in pristine and impacted Mediterranean seagrass sediments. *Aquatic Microbial Ecology* 36(3):227–237.
- Holmer, M., N. Marba, E. Diaz-Almela, C.M. Duarte, M. Tsapakis, and R. Danavaro. 2007. Sedimentation of organic matter from fish farms in oligotrophic Mediterranean assessed through bulk and stable isotope (δ13C and δ15N) analyses. *Aquaculture* 262:268–280.
- Kennedy, H., E. Gacia, D.P. Kennedy, S. Papadimitriou, and C.M. Duarte. 2004. Organic carbon sources to SE Asian coastal sediments. *Estuarine Coastal and Shelf Science* 60(1):59–68.
- Kennedy, H., J. Beggins, C.M. Duarte, J.W. Fourqurean, M. Holmer, N. Marba, and J.J. Middelburg. 2010. Seagrass sediments as a global carbon sink: Isotopic constraints. *Global Biogeochemical Cycles* 24(GB4026):1–8.
- Kenworthy, J. and G. Thayer. 1984. Production and decomposition of the roots and rhizomes of seagrasses, Zostera marina and Thalassia testudinum, in temperate and subtropical marine ecosystems. *Bulletin of Marine Science* 35(364–379).
- Koch, M.S. and C.J. Madden. 2001. Patterns of primary production and nutrient availability in a Bahamas lagoon with fringing mangroves. *Marine Ecology Progress Series* 219:109–119.
- Lee-Nagel, J. 2007. Plant-sediment interaction and biogeochemical cycling for seagrass communities in the Chesapeake and Florida bays. University of Maryland.

- Lindeboom, H.J. and J.J. Sandee. 1989. Production and consumption of tropical seagrass fields in eastern Indonesia measured with bell jars and microelectrodes. *Netherlands Journal of Sea Research* 23(2):181–190.
- Lo Iacono, C., M.A. Mateo, E. Gracia, L. Guasch, R. Carbonell, L. Serrano, O. Serrano, and J. Danobeita. 2008. Very high-resolution seismo-acoustic imaging of seagrass meadows (Mediterranean Sea): Implications for carbon sink estimates. *Geophysical Research Letters* 35:L18601.
- Martin, S., J. Clavier, J.-M. Guarini, and L. Chauvaud. 2005. Comparison of Zostera marina and maerl community metabolism. *Aquatic Botany* 83:161–174.
- Mateo, M.A., J. Romero, M. Perez, M.M. Littler, and D.S. Littler. 1997. Dynamics of millenary organic deposits resulting from the growth of the mediterranean seagrass *Posidonia oceanica*. *Estuarine Coastal and Shelf Science* 44:103–110.
- Moncreiff, C.A., M.J. Sullivan, and A.E. Daehnick. 1992. Primary production dynamics in seagrass beds of the Mississippi Sound: The contributions of seagrass, epiphytic algae, sand microflora, and phytoplankton. *Marine Ecological Progress Series* 87:161–171.
- Morgan, M.D. and C.L. Kitting. 1984. Productivity and utilization of the seagrass Halodule wrightii and its attached epiphytes. *Limnology and Oceanography* 29(5):1066–1076.
- Moriarty, D.J.W., D.G. Roberts, and P.C. Pollard. 1990. Primary and bacterial productivity of tropical seagrass communities in the Gulf of Carpentaria, Australia. *Marine Ecology Progress Series* 61:145–157.
- Murray, B.C., L. Pendleton, W.A. Jenkins, and S.D. Sifleet. 2011. Green payments for blue carbon: Economic incentives for protecting threatened coastal habitats. Durham, NC: Nicholas Institute for Environmental Policy Solutions, Duke University.
- Murray, L. and R.L. Wetzel. 1987. Oxygen production and consumption associated with the major autotrophic components in two temperate seagrass communities. *Marine Ecology Progress Series* 38:231–239.
- Nixon, S.W. and C.A. Oviatt. 1972. Preliminary measurements of midsummer metabolism in beds of eelgrass, Zostera marina. *Ecology* 53(150–153).
- Odum, H.T. 1956. Primary production in flowing waters. Limnology and Oceanography 1:102-117.

———. 1959. Measurements of productivity of turtle grass flats reefs and the Bahia Fosforescente of southern Puerto Rico. *Institute of Marine Science of the University of Texas* 6:159–170.

- ———. 1962. Further studies on reaeration and metabolism of Texas Bays 1958–1960. *Institute of Marine Science of the University of Texas* 8:23–55.
- Pasqualani, V., C. Pergent-Martini, P. Clabaut, and G. Pergent. 1998. Mapping of *Posidonia oceanica* using aerial photographs and side scan sonar: Application off the island of Corsica (France). *Estuarine Coastal and Shelf Science* 47:359–367.
- Pellikaan, G. and P. Nienhuis. 1988. Nutrient uptake and release during growth and decomposition of eelgrass, Zostera marina L., and its effect on the nutrien dynamics of Lake Grevelingen. *Aquatic Botany* 30:189–214.
- Penhale, P.A. 1977. Macrophyte-epiphyte biomass and productivity in an eelgrass (Zostera marina L.) community. *Journal of Experimental Marine Biology and Ecology* 26(2):211–224.
- Plus, M., J.-M. Deslous-Paoli, I. Auby, and F. Dagault. 2001. Factors influencing primary production of seagrass beds (Zostera noltii Hornem) in the Thau lagoon (French Mediterranean coast). *Journal of Experimental Marine Biology and Ecology* 259:62–84.
- Qasim, S.Z. and P.M.A. Bhattathiri. 1971. Primary production of a seagrass bed on Kavaratti Atoll (Laccadives). *Hydrobiologia* 38:29–38.
- Reyes, E. and M. Merino. 1991. Diel dissolved oxygen dynamics and eutrophication in a shallow well-mixed tropical lagoon (Cancun, Mexico). *Estuaries* 14(4):372–381.
- Risgaard-Petersen, N. and L.D.M. Ottosen. 2000. Nitrogen cycling in two temperate Zostera marina beds:seasonal variation. *Marine Ecology Progress Series* 198:93–107.

- Romero, J., M. Perez, M.A. Mateo, and E. Sala. 1994. The belowground organs of the Mediterranean seagrass *Posidonia oceanica* as a biogeochemical sink. *Aquatic Botany* 47:13–19.
- Stutes, J., J. Cebrian, A.L. Stutes, A. Hunter, and A.A. Corcoran. 2007. Benthic metabolism across a gradient of anthropogenic impact in three shallow coastal lagoons in NW Florida. *Marine Ecological Progress Series* 348:55– 70.
- Viaroli, P., M. Bartoli, C. Bondavalli, R.R. Christian, G. Giordani, and M. Naldi. 1996. Macrophyte communities and their impact on benthic fluxes of oxygen, sulphide and nutrients in shallow eutrophic environments. *Hydrobiologia* 329:105–119.
- Waycott, M., C.M. Duarte, T.J.B. Carruthers, R.J. Orth, W.C. Dennison, S. Olyarnik, A. Calladine, J.W. Fourqurean, K.L. Heck, A.R. Hughes, G.A. Kendrick, W.J. Kenworthy, F.T. Short, and S.L. Williams. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proceedings of the National Academy of Sciences of the United States of America* 106(30):12377–12381.
- Welsh, D.T., M. Bartoli, D. Nizzoli, G. Castaldelli, S.A. Riou, and P. Viaroli. 2000. Denitrification, nitrogen fixation, community primary productivity and inorganic-N and oxygen fluxes in an intertidal Zostera noltii meadow. *Marine Ecology Progress Series* 208:65–77.
- Wium-Andersen, S. and J. Borum. 1984. Biomass variation and autotrophic production of an epiphyte-macrophyte community in a coastal Danish area: Eelgrass (Zostera marina L.) biomass and net production. *Ophelia* 23:33–46.
- Yarbro, L.A. and P.R.J. Carlson. 2008. Community oxygen and nutrient fluxes in seagrass beds of Florida Bay, USA. *Estuaries and Coasts* 31:877–897.
- Ziegler, S. and R. Benner. 1999. Nutrient cycling in the water column of a subtropical seagrass meadow. *Marine Ecology Progress Series* 188:51–62.

References

- 1. Macintyre, I.G., M.M. Littler, and D.S. Littler. 1995. Holocene history of Tobacco Range, Belize, Central America. *Atoll Research Bulletin* 430:1–18.
- 2. Donato, D., et al. 2011. Mangroves among the most carbon-rich forests in the tropics. *Nature Geoscience Online*: April 3.
- 3. Cebrian, J. 2002. Variability and control of carbon consumption, export, and accumulation in marine communities. *Limnology and Oceanography* 47(1):11–22.
- 4. Chmura, G.L., et al. 2003. Global carbon sequestration in tidal, saline wetland soils. *Global Biogeochemical Cycles* 17(4):1–12.
- 5. Fujimoto, K., et al. 1991. Belowground carbon storage of Micronesian mangrove forests. *Ecological Research* 14: 409–413.
- 6. Nellemann, C., et al. 2009. *Blue carbon*. A rapid response assessment. United Nations Environment Programme.
- 7. Duarte, C.M., J.J. Middleburg, and N. Caraco. 2005. Major role of marine vegetation on the oceanic carbon cycle. *Biogeosciences* 2:1–8.
- 8. Jennerjahn, T.C., and V. Ittekkot. 2002. Relevance of mangroves for the production and deposition of organic matter along tropical continental margins. *Naturwissenschaften* 89:23–30.
- 9. Bouillion, S., et al. 2009. Mangroves. In *The management of natural coastal carbon sinks*, edited by D.A. Laffoley and G. Grimsditch, 13–20. Gland Switzerland: IUCN.
- 10. Twilley, R.R., R.H. Chen, and T. Hargis. 1992. Carbon sinks in mangroves and their implications for carbon budget of tropical ecosystems. *Water, Air, and Soil Pollution* 63:265–288.
- 11. Bridgham, S.D., et al. 2006. The carbon balance of North American wetlands. *Wetlands* 26(4):889–916.
- 12. Vegas-Vilarrubia, T., et al. 2010. Tropical histosols of the lower Orinoco delta, features and preliminary quantification of their carbon storage. *Geoderma* 155:280–288.
- 13. PWA and SAIC. 2009. Greenhouse gas mitigation typology issues paper. Tidal Wetlands Restoration. California Climate Action Registry.

- 14. Bouillion, S., et al. 2008. Mangrove production and carbon sinks: A revision of global budget estimates. *Global Biogeochemical Cycles* 22:GB2013.
- 15. FAO (Food and Agriculture Organization). 2007. The World's Mangroves 1980–2005. Rome: FAO.
- 16. UNEP (United Nations Environment Programme) and FAO. 1981a. Proyecto de evaluacion de los recursos forestales tropicales: Los recursos forestales de la America tropical. Rome: UNEP and FAO.
- 17. ———. 1981b. Tropical forest resources assessment project: Forest resources of tropical Africa. Part II. Country briefs. Rome: UNEP and FAO.
- 18. ———. 1981c. Tropical forest resources assessment project: Forest resources of tropical Asia. Rome: UNEP and FAO.
- 19. FAO. 2001. Global forest resources assessment 2000: Main report. FAO Forestry Paper 140. Rome: FAO.
- 20. FAO. 2006. Global forest resources assessment 2005: Progress toward sustainable forest management. FAO Forestry Paper 147. Rome: FAO.
- 21. Murray, B.C., et al. 2011. Green payments for blue carbon: Economic incentives for protecting threatened coastal habitats. Durham, NC: Nicholas Institute for Environmental Policy Solutions, Duke University.
- 22. Craft, C.B. and C.J. Richardson. 1998. Recent and long-term organic soil accretion and nutrient accumulation in the Everglades. *Soil Science Society of America Journal* 62:834–843.
- 23. Hussein, A.H., M.C. Rabenhorst, and M.L. Tucker. 2004. Modeling of carbon sequestration in coastal marsh soils. *Soil Science Society of America Journal* 68:1786–1795.
- 24. Johnson, B.J., et al. 2007. Middle to late Holocene fluctuations of C3 and C4 vegetation in a northern New England salt marsh, Sprague Marsh, Phippsburg, Maine. *Organic Geochemistry* 38: 398–403.
- 25. Choi, Y. and Y. Wang. 2004. Dynamics of carbon sequestration in a coastal wetland using radiocarbon measurements. *Global Biogeochemical Cycles* 18:GB4016.
- 26. Brevik, E.C. and J.A. Homburg. 2004. A 5000-year record of carbon sequestration from a coastal lagoon and wetland complex, Southern California, USA. *Catena* 57: 221–232.
- 27. Giani, L., et al. 1996. Methanogenesis in saltmarsh soils of the North Sea coast of Germany. European Journal of Soil Science 47: 175–182.
- 28. Drexler, J.Z., C.S. de Fontaine, and S.J. Deverel. 2009. The legacy of wetland drainage on the remaining peat in the Sacramento San Joaquin Delta, California, USA. *Wetlands* 29(1): 372–386.
- 29. Choi, Y. and Y. Wang. 2001. Vegetation succession and carbon sequestration in a coastal wetland in northwest Florida: Evidence from carbon isotopes. *Global Biogeochemical Cycles* 15(2):311–319.
- 30. Craft, C.B., S.W. Broome, and E.D. Seneca. 1988. Nitrogen, phosphorus, and organic carbon pools in natural and transplanted marsh soils. *Estuaries* 11(4):272–280.
- 31. Hemminga, M.A., et al. 1996. Assessment of carbon allocation and biomass production in a natural stand of the salt marsh plant *Spartina anglica* using 13C. *Marine Ecology Progress Series* 130:169–178.
- 32. Yu, O.T. and G.L. Chmura. 2009. Soil carbon may be maintained under grazing in a St. Lawrence estuary tidal marsh. *Environmental Conservation* 36(4):312–320.
- 33. Morgan, P.A. and F.T. Short. 2002. Using functional trajectories to track constructed salt marsh development in the Great Bay estuary, Maine/New Hampshire U.S.A. *Restoration Ecology* 10(3):461–473.
- 34. Morris, J.T. and B. Haskin. 1990. A 5-year record of areal primary production and stand characteristics of *Spartina alterniflora. Ecology* 71(6):2209–2217.
- 35. Teal, J.M. and B.L. Howes. 1996. Interannual variability of a salt-marsh ecosystem. *Limnology and Oceanography* 41(4):802–809.
- 36. Yang, S. and J. Chen. 1995. Coastal salt marshes and mangrove swamps in China. Chinese *Journal of Oceanology and Limnology* 13(4):318–324.
- 37. Adam, P. 2002. Saltmarshes in a time of change. Environmental Conservation 29(1):39-61.

- 38. Duarte, C.M., et al. 2008. The charisma of coastal ecosystems: Addressing the imbalance. *Estuaries and Coasts* 31:233–238.
- 39. Huang, Y., et al. 2009. Marshland conversion to cropland in northeast China from 1950 to 2000 reduced the greenhouse effect. *Global Change Biology* 16(2):680–695.
- 40. Armentano, T.V. and E.S. Menges. 1986. Patterns of change in the carbon balance of organic soil wetlands of the temperate zone. *Journal of Ecology* 74(3):755–774.
- 41. Ewing, J.M. and M.J. Vepraskas. 2006. Estimating primary and secondary subsidence in an organic soil 15, 20, and 30 years after drainage. *Wetlands* 26(1):119–130.
- 42. Duarte, C.M., et al. 2101. Seagrass community metabolism: Assessing the carbon sink capacity of seagrass meadows. *Global Biogeochemical Cycles* 24(GB4032):1–8. Mateo, M.A., et al., Dynamics of millenary organic deposits resulting from the growth of the mediterranean seagrass *Posidonia oceanica. Estuarine Coastal and Shelf Science* 44:103–110.
- 43. Romero, J., et al. 1994. The belowground organs of the Mediterranean seagrass *Posidonia oceanica* as a biogeochemical sink. *Aquatic Botany* 47:13–19.
- 44. Lo Iacono, C., et al. 2008. Very high-resolution seismo-acoustic imaging of seagrass meadows (Mediterranean Sea): Implications for carbon sink estimates. *Geophysical Research Letters* 35:L18601.
- 45. Kennedy, H., et al. 2010. Seagrass sediments as a global carbon sink: Isotopic constraints. *Global Biogeochemical Cycles* 24(GB4026):8.
- 46. Bouillon, S. and H.T.S. Boschker. 2006. Bacterial carbon sources in coastal sediments: A cross-system analysis based on stable isotope data of biomarkers. *Biogeosciences* 3:175–185.
- 47. Duarte, C.M. and C.L. Chiscano. 1999. Seagrass biomass and production: A reassessment. *Aquatic Botany* 65:159–174.
- 48. Waycott, M., et al. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. *Proceedings of the National Academy of Sciences of the United States of America* 106(30):12377–12381.
- 49. Pasqualani, V., et al. 1998. Mapping of *Posidonia oceanica* using aerial photographs and side scan sonar: Application off the island of Corsica (France). *Estuarine Coastal and Shelf Science* 47:359–367.
- 50. Charpy-Roubaud, C. and A. Sournia. 1990. The comparative estimation of phytoplankton microphytobenthic production in the oceans. *Marine Microbial Food Webs* 4:31–57.
- 51. Cahoon, D.R. and J.C. Lynch. 1997. Vertical accretion and shallow subsidence in a mangrove forest of southwestern Florida. *Mangroves and Salt Marshes* 1:173–186.
- 52. Callaway, J.C., R.D. DeLuane, and W.H.J. Patrick. 1997. Sediment accretion rates from four coastal wetlands along the Gulf of Mexico. *Journal of Coastal Research* 13:181–191.
- 53. Lynch, J.C. 1989. Sedimentation and nutrient accumulation in mangrove ecosystems of the Gulf of Mexico. Lafayette, LA: University of Southwestern Lousiana.
- 54. Twilley, R., A. Lugo, and C. Patterson-Zuca. 1986. Litter production and turnover in basin mangrove forests in southwest Florida. *Ecology* 67:670–683.
- 55. Brunskill, G.J., J. Zagorskis, and J. Pfitzner. 2002. Carbon burial rates in sediments and a carbon mass balance, of the Herbert River region of the Great Barrier Reef continental shelf, north Queensland, Australia. *Estuarine Coastal and Shelf Science* 54:677–700.
- 56. Alongi, D.M., et al. 1999. Sources, sinks, and export of organic carbon fluzes in adjacent tropical nearshore sediments. *Marine Ecological Progress Series* 56:134–144.
- 57. Robertson, A. and P. Daniel. 1989. Decomposition and the annual flux of detritus from fallen timber in tropical mangrove forests. *Limnology and Oceanography* 34:640–647.
- Lee, S. 1990. Net areal primary productivity, litter production and decomposition of the reed Phragmites communis in a nature reserve in Hong Kong: Management implications. *Marine Ecology Progress Series* 66:161– 173.

- 59. Li, M.S. and S.Y. Lee. 1998. Carbon dynamics of Deep Bay, eastern Pearl River Estuary, China: A mass balance budget and implications for shorebird conservation. *Marine Ecology Progress Series* 172:73–87.
- 60. Van der Valk, A. and P. Attiwill. 1986. Decomposition of leaf and root litter of Aviccenia marina at Westernport Bay, Victoria, Autralia. *Aquatic Botany* 18:205–221.
- 61. Ong, J.E. 1993. Mangroves: A carbon source and sink. Chemosphere 27:1097–1107.
- 62. Golley, F., H.T. Odum, and R. Wilson. 1962. The structure and metabolism of a Puerto Rican red mangrove forest in May. *Ecology* 43:8–19.
- 63. Emmerson, W.D. and L.E. McGwynne. 1992. Feeding and assimilation of mangrove leaves by the crab Sesarma meinerti de Man in relation to leaf-litter production in Mgazana, a warm-temperate southern African mangrove swamp. *Journal of Experimental Marine Biology and Ecology* 157(1):41–53.
- 64. Chen, R.H. and R.R. Twilley. 1999. A simulation model of organic matter and nutrient accumulation in mangrove wetland soils. *Biogeochemistry* 44(1):93–118.
- 65. Ross, M.S., et al. 2000. The southeast saline Everglades revisited: 50 years of coastal vegetation change. *Journal of Vegetation Science* 11:101–112.
- 66. Naidoo, G. 1980. Mangrove soils of the Beachwood area, Durban. Journal of South African Botany 46:293-304.
- 67. Cardona, P. and L. Botero. 1998. Soil characteristics and vegetation structure in a heavily deteriorated mangrove forest in the Caribbean coast of Colombia. *Biotropica* 30:24–34.
- 68. Fujimoto, K., et al. 1996. Mangrove habitat formation and response to holocene sea-level changes on Krosae Island, Micronesia. *Mangroves and Salt Marshes* 1:47–57.
- 69. Fujimoto, K. and T. Miyagi. 1993. Development process of tidal-flat type mangrove habitats and their zonation in the Pacific Ocean. *Vegetatio* 106:137–146.
- 70. Day, J., et al. 1987. The productivity and composition of mangrove forests, Laguna de Terminos, Mexico. *Aquatic Botany* 27:267–284.
- 71. Lin, P., et al. 1985. The biomass and productivity of Kandelia camel community. *Journal of Xiamen University* 14:508–514.
- 72. Lin, P., et al. 1990. Biomass and productivity of Bruguiera sexanguia mangrove forest in Hainan Island, China. *Journal of Xiamen University* 29:209–213.
- 73. Lee, S.Y. 1990. Primary productivity and particulate organic matter flow in an estuarine mangrove-wetland in Hong Kong. *Marine Biology* 106:453–463.
- 74. Christensen, B. 1978. Biomass and primary production of Rhizophora apiculata B1. in a mangrove forest in southern Thailand. *Aquatic Botany* 4:43–52.
- 75. Cahoon, D.R. 1994. Recent accretion in two managed marsh impoundments in coastal Louisiana. Ecological Applications 41:166–176.
- 76. Cahoon, D.R. and R.E. Turner. 1989. Accretion and canal impacts in a rapidly subsiding wetland: Feldspar marker horizon technique. *Estuaries* 12:260–268.
- 77. Hatton, R.S. 1981. Aspects of marsh accretion and geochemistry: Barataria Basin. Baton Rouge, LA: Lousiana State University.
- 78. Bryant, J.C. and R.H. Chabreck. 1998. Effects of impoundment on vertical accretion of coastal marsh. *Estuaries* 21:416–422.
- 79. Markewich, H.W. 1998. Carbon storage and late Holocene chronostratigraphy of a Mississippi River deltaic marsh. U.S. Geological Survey Open File Report 98–36.
- 80. Day, J., et al. 1973. Community structure and carbon budget of a salt marsh and shallow bay estuarine system in Louisiana. Baton Rouge, LA: Center for Wetland Resources, Louisana State University.
- 81. Hopkinson, O., J.G. Gosselink, and P. Panardo. 1978. Aboveground production of seven marsh plant species in coastal Lousiana. *Ecology* 59:760–769.

- 82. White, D., et al. 1978. Production and decomposition of the dominant salt marsh plants in Lousiana. *Ecology* 59:751–759.
- 83. Connor, R.F., G. Chmura, and C.B. Beecher. 2001. Carbon accumulation in Bay of Fundy salt marshes: Implications for restoration of reclaimed marshes. *Global Biogeochemical Cycles* 15(4):934–954.
- 84. Chmura, G.L. and G. Hung. 2004. Controls on salt marsh accretion: A test in salt marshes of Eastern Canada. *Estuaries* 27(1):70–81.
- 85. Anisfield, S.C., M.J. Tobin, and G. Benoit. 1999. Sedimentation rates in flow-restricted and restored salt marshes in Long Island Sound. *Estuaries* 22:231–244.
- 86. Roman, C.T., et al. 1997. Accretion of a New England (U.S.A.) salt marsh in response to inlet migration, storms and sea-level rise. *Estuarine Coastal and Shelf Science* 45:717–727.
- 87. McCaffery, R.J. and J. Thomson. 1980. A record of the accumulation of sediment trace metals in a Connecticut salt marsh. *Advances in Geophysics* 22:165–236.
- 88. Orson, R.A., R.S. Warren, and W.A. Niering. 1998. Interpreting sea-level rise and rates of vertical marsh accretion in a southern New England tidal salt marsh. *Estuarine Coastal and Shelf Science* 26:419–429.
- 89. Howes, B.L., J. Dacey, and J.M. Teal. 1985. Annual C mineralization and belowground production of *Spartina alterniflora* in a New England salt marsh. *Ecology* 66:595–605.
- 90. Kearney, M.S. and J.C. Stevenson. 1991. Island land loss and marsh vertical accretion rate evidence for historical sea-level changes in the Chesapeake Bay. *Journal of Coastal Research* 7:403–415.
- 91. Blum, L. 1993. *Spartina alterniflora* root dynamics in a Virginia marsh. *Marine Ecology Progress Series* 102:169–178.
- 92. Cahoon, D.R. and J.C. Stevenson. 1986. Production, predation and decomposition in a low-salinity Hibischus marsh. *Ecology* 67:1341–1350.
- 93. Callaway, J.C., R.D. DeLuane, and W.H.J. Patrick. 1996. Chernobyl 137Cs used to determine sediment accretion rates at selected northern European salt marshes. *Limnology and Oceanography* 41:444–450.
- 94. Oenema, O. and R.D. DeLuane. 1988. Accretion rates in salt marshes in the Eastern Scheldt, southwest Netherlands. *Estuarine Coastal and Shelf Science* 26:379–394.
- 95. Buth, G. 1987. Decomposition of roots of three plant communities in a Dutch salt marsh. *Aquatic Botany* 29:123–128.
- 96. French, J.R. and T. Spencer. 1993. Dynamics of sedimentation in a tide-dominated back barrier salt marsh, Norfolk, UK. Marine Geology 110:315–331.
- 97. Cahoon, D.R., J.C. Lynch, and A. Powell. 1996. Marsh vertical accretion in a southern California estuary, U.S.A. *Estuarine Coastal and Shelf Science* 43:19–36.
- 98. Patrick, W.H.J. and R.D. DeLuane. 1990. Subsidence, accretion, and sea-level rise in south San Fransisco Bay marshes. *Limnology and Oceanography* 35:1389–1395.
- 99. Craft, C.B., E.D. Seneca, and S.W. Broome. 1993. Vertical accretion in micro-tidal regularly and irregularly flooded estuarine marshes. *Estuarine Coastal and Shelf Science* 37:371–386.
- 100. Hensel, P.F., J.W.J. Day, and D. Pont. 1999. Wetland vertical accretion and soil elevation change in the Rhone River Delta, France: The importance of riverine flooding. *Journal of Coastal Research* 15:668–681.
- 101. Teal, J.M. 1962. Energy flow in the salt marsh ecosystem of Georgia. Ecology 43:615–624.
- 102. Morris, J.T. and A. Jensen. 1998. The carbon balance of grazed and non-grazed *Spartina anglica* salt marshes at Skalligen, Denmark. *Journal of Ecology* 86:229–242.
- 103. Lee-Nagel, J. 2007. Plant-sediment interaction and biogeochemical cycling for seagrass communities in the Chesapeake and Florida Bays. University of Maryland.
- 104. Yarbro, L.A. and P.R.J. Carlson. 2008. Community oxygen and nutrient fluxes in seagrass beds of Florida Bay, USA. *Estuaries and Coasts* 31:877–897.

- 105. Martin, S., et al. 2005. Comparison of Zostera marina and maerl community metabolism. *Aquatic Botany* 83:161–174.
- 106. Odum, H.T. 1956. Primary production in flowing waters. Limnology and Oceanography 1:102-117.
- 107. Kenworthy, J. and G. Thayer. 1984. Production and decomposition of the roots and rhizomes of seagrasses, Zostera marina and Thalassia testudinum, in temperate and subtropical marine ecosystems. *Bulletin of Marine Science* 35:364–379.
- 108. Barron, C., et al. 2004. Community metabolism and carbon budget along a gradient of seagrass (*Cymodocea nodosa*) colonization. *Limnology and Oceanography* 49:1642–1652.
- 109. Gazeau, F., et al. 2005. Whole-system metabolism and CO₂ fluxes in a Mediterranean Bay dominated by seagrass beds (Palma Bay, NW Mediterranean). *Biogeosciences* 2:43–60
- 110. Holmer, M., et al. 2004. Carbon cycling and bacterial carbon sources in pristine and impacted Mediterranean seagrass sediments. *Aquatic Microbial Ecology* 36(3):227–237.
- 111. Cebrian, J., et al. 2000. Fate and production of the seagrass *Cymodocea nodosa* in different stages of meadow formation. *Marine Ecology Progress Series* 204:119–130.
- 112. Odum, H.T. 1963. Productivity measurements in Texas turtle grass and the effects of dredging an intracoastal channel. *Institute of Marine Science of the University of Texas* 6:48–58.
- 113. Stutes, J., et al. 2007. Benthic metabolism across a gradient of anthropogenic impact in three shallow coastal lagoons in NW Florida. *Marine Ecological Progress Series* 348:55–70.
- 114. Herbert, D.A. and J.W. Fourqurean. 2008. Ecosystem structure and function still altered two decades after short-term fertilization of a seagrass meadow. *Ecosystems* 11:688–700.
- 115. Calleja, M.L., et al. 2006. Light regulation of benthic sulfate reduction rates mediated by seagrass (Thalassia testudinum) metabolism. *Estuaries and Coasts* 29:1255–1264.
- 116. Risgaard-Petersen, N. and L.D.M. Ottosen. 2000. Nitrogen cycling in two temperate Zostera marina beds: Seasonal variation. *Marine Ecology Progress Series* 198:93–107.
- 117. Plus, M., et al. 2001. Factors influencing primary production of seagrass beds (Zostera noltii Hornem) in the Thau lagoon (French Mediterranean coast). *Journal of Experimental Marine Biology and Ecology* 259:62–84.
- 118. Viaroli, P., et al. 1996. Macrophyte communities and their impact on benthic fluxes of oxygen, sulphide and nutrients in shallow eutrophic environments. *Hydrobiologia* 329:105–119.
- 119. Frankingnoulle, M. and J.M. Bouquegneau. 1987. Seasonal variation of the diel carbon budget of a marine macrophyte ecosystem. *Marine Ecology Progress Series* 38:197–199.
- 120. Welsh, D.T., et al. 2000. Denitrification, nitrogen fixation, community primary productivity and inorganic-N and oxygen fluxes in an intertidal Zostera noltii meadow. *Marine Ecology Progress Series* 208:65–77.
- 121. Reyes, E. and M. Merino. 1991. Diel dissolved oxygen dynamics and eutrophication in a shallow well-mixed tropical lagoon (Cancun, Mexico). *Estuaries* 14(4):372–381.
- 122. Murray, L. and R.L. Wetzel. 1987. Oxygen production and consumption associated with the major autotrophic components in two temperate seagrass communities. *Marine Ecology Progress Series* 38:231–239.
- 123. Caffrey, J.M. 2004. Factors controlling net ecosystem metabolism in U.S. estuaries. Estuaries 27:90-101.
- 124. Anton, A., et al. 2009. Low impact of Hurricane Katrina on seagrass community structure and functioning in the northern Gulf of Mexico. *Bulletin of Marine Science* 85(1):45–59.
- 125. Odum, H.T. 1962. Further studies on reaeration and metabolism of Texas Bays 1958–1960. *Institute of Marine Science of the University of Texas* 8:23–55.
- 126. Ziegler, S. and R. Benner. 1999. Nutrient cycling in the water column of a subtropical seagrass meadow. *Marine Ecology Progress Series* 188:51–62.
- 127. Moriarty, D.J.W., D.G. Roberts, and P.C. Pollard. 1990. Primary and bacterial productivity of tropical seagrass communities in the Gulf of Carpentaria, Australia. *Marine Ecology Progress Series* 61:145–157.

- 128. Erftemeijer, P.L.A., R. Osinga, and A.E. Mars. 1993. Primary production of seagrass beds in South Sulawesi (Indonesia): A comparison of habitats, methods and species. *Aquatic Botany* 46:67–90.
- 129. Apostolaki, E.T., et al. 2010. Metabolic imbalance in coastal vegetated (*Posidonia oceanica*) and unvegetated benthic ecosystems. *Ecosystems* 13:459–471.
- 130. Gacia, E., et al. 2005. Light-dependence of the metabolic balance of a highly productive Philippine seagrass community. *Journal of Experimental Marine Biology and Ecology* 316:55–67.
- 131. Eyre, B.D. and J.P. Ferguson. 2002. Comparison of carbon production and decomposition, benthic nutrient fluxes and denitrification in seagrass, phytoplankton, benthic microalgae- and macroalgae-dominated warm-temperate Australian lagoons. *Marine Ecology Progress Series* 229:43–59.
- 132. Koch, M.S. and C.J. Madden. 2001. Patterns of primary production and nutrient availability in a Bahamas lagoon with fringing mangroves. *Marine Ecology Progress Series* 219:109–119.
- 133. Duarte, C.M., R. Martinez, and C. Barron. 2002. Biomass, production and rhizome growth near the northern limit of seagrass (Zostera marina) distribution. *Aquatic Botany* 72:183–189.
- 134. Odum, H.T. 1959. Measurements of productivity of turtle grass flats reefs and the Bahia Fosforescente of southern Puerto Rico. *Institute of Marine Science of the University of Texas* 6:159–170.
- 135. Qasim, S.Z. and P.M.A. Bhattathiri. 1971. Primary production of a seagrass bed on Kavaratti Atoll (Laccadives). *Hydrobiologia* 38:29–38.
- 136. Baird, D. and R.E. Ulanowicz. 1993. Comparative study on the trophic structure, cycling and ecosystem properties of four tidal estuaries. *Marine Ecology Progress Series* 99:221–237.
- 137. Asmus, R.M., M. Sprung, and H. Asmus. 2000. Nutrient fluxes in intertidal communities of a South European lagoon (Ria Formos): Similarities and differences with a northern Wadden Sea bay (Sylt-Romo Bay). *Hydrobiologia* 436:217–235.
- 138. Pellikaan, G. and P. Nienhuis. 1988. Nutrient uptake and release during growth and decomposition of eelgrass, Zostera marina L., and its effect on the nutrien dynamics of Lake Grevelingen. *Aquatic Botany* 30:189–214.
- 139. Fourqurean, J.W., M.F. Muth, and J.N. Boyer. 2010. Epiphyte loads on seagrasses and microphytobenthos abundance are not reliable indicators of nutrient availability in coastal ecosystems. *Marine Pollution Bulletin* 60:971–983.
- 140. Fourqurean, J.W., et al. 2001. Spatial and temporal pattern in seagrass community composition and productivity in south Florida. *Marine Biology* 138:341–354.
- 141. Fourqurean, J.W., et al. 2005. Spatial and seasonal variability in elemental content, d13C and d15N of Thalassia testudinum from south Florida. *Estuaries* 28(3): 447–461.
- 142. Armitage, A.R. and J.W. Fourqurean. 2009. Stable isotopes reveal complex changes in trophic relationships following nutrient addition in a coastal marine ecosystem. *Estuaries and Coasts* 32:1152–1164.
- 143. Barron, C. and C.M. Duarte. 2009. Dissolved organic matter (DOM) release in a *Posidonia oceanica* meadow. *Marine Ecological Progress Series* 374:75–84.
- 144. Heffernan, J.J. and R.A. Gibson. 1983. A comparison of primary production rates in Indian River, Florida seagrass systems. *Florida Science* 48:295–306.
- 145. Wium-Andersen, S. and J. Borum. 1984. Biomass variation and autotrophic production of an epiphytemacrophyte community in a coastal Danish area: Eelgrass (Zostera marina L.) biomass and net production. *Ophelia*, 23:33–46.
- 146. Lindeboom, H.J. and J.J. Sandee. 1989. Production and consumption of tropical seagrass fields in eastern Indonesia measured with bell jars and microelectrodes. *Netherlands Journal of Sea Research* 23(2):181–190.
- 147. Moncreiff, C.A., M.J. Sullivan, and A.E. Daehnick. 1992. Primary production dynamics in seagrass beds of the Mississippi Sound: The contributions of seagrass, epiphytic algae, sand microflora, and phytoplankton. *Marine Ecological Progress Series* 87:161–171.
- 148. Morgan, M.D. and C.L. Kitting. 1984. Productivity and utilization of the seagrass Halodule wrightii and its attached epiphytes. *Limnology and Oceanography* 29(5):1066–1076.

- 149. Holmer, M., et al. 2007. Sedimentation of organic matter from fish farms in oligotrophic Mediterranean assessed through bulk and stable isotope (δ13C and δ15N) analyses. *Aquaculture* 262: 268–280.
- 150. Kennedy, H., et al. 2004. Organic carbon sources to SE Asian coastal sediments. *Estuarine Coastal and Shelf Science* 60(1):59–68.
- 151. Holmer, M., et al. 2001. The importance of sulfate reduction based mineralization for nutrient regeneration in tropical seagrass sediments. *Aquatic Botany* 71:1–17.
- 152. Nixon, S.W. and C.A. Oviatt. 1972. Preliminary measurements of midsummer metabolism in beds of eelgrass, Zostera marina. *Ecology* 53:150–153.
- 153. Penhale, P.A. 1977. Macrophyte-epiphyte biomass and productivity in an eelgrass (Zostera marina L.) community. *Journal of Experimental Marine Biology and Ecology* 26(2): 211–224.



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